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4-Aminoquinoline-hybridization *en route* towards the development of rationally designed antimalarial agents

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The resistance of *Plasmodium falciparum*, the causative agent of malaria, against quinine and chloroquine along with the lack of malaria vaccines has encouraged the development of various synthetic strategies towards biologically active scaffolds. An emerging strategy in medicinal chemistry, termed molecular hybridization, involves the covalent fusion of two or more drugs, active compounds, and/or pharmacophoric units into a hybrid compound, with fascinating activities and multiple but not essentially simultaneous pharmacological targets. 4-Aminoquinolines are considered as promising antimalarials and 4-aminoquinoline hybridization is considered as an attractive and feasible approach for the development of new molecular frameworks for averting and delaying the emergence of drug resistance along with improved efficacy. The present review article describes the recent developments on the 4-aminoquinoline-hybridization towards the development of new antimalarials.

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1. Introduction

In the 20th century, the drug design approach accomplished the “one-target-one-drug” concept for the discovery of new drugs which, without any iota of doubt, will be dominant for many years. According to this concept, a single drug is designed for a



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extensively on the synthesis and antimalarial evaluation of 4-aminoquinoline based molecular conjugates. He has published sixteen research papers in international journals of repute and is currently working as an Assistant Professor at Khalsa College, Amritsar. His research interests include synthesis of novel molecular frameworks targeting tropical infections. He is also engaged in utilizing β -lactam synthon methodology for the preparation of functionally enriched heterocyclic scaffolds with biological relevance.



Dr Kirkwood Land is an Associate Professor in the Department of Biological Sciences at the University of the Pacific. Dr Land's research group explore interdisciplinary approaches to drug discovery. In particular, he is interested in strategies for the control of microorganisms, especially, bacteria and parasites associated with domesticated animals and humans. His work utilizes the One Health

approach, which integrates an understanding of humans, animals, and the environment, to address current problems of infectious diseases.

single target. Although numerous drugs have been designed and used clinically with good success using this approach, the strategy of one drug hammering one target remains inadequate for the treatment of several diseases, such as neurodegenerative syndromes, cardiovascular diseases, diabetes and cancer, which involve multiple pathogenic factors.¹ Further, an ideal drug for one target does not always have clinical efficacy due to non-recognition of the *in vivo* target or the inability to access the site of action. This target-based strategy does not always guarantee success since some selective drugs can work only in a selected number of patients. For example, Astra Zeneca's Iressa (gefitinib) is designed to treat lung cancer *via* targeting EGFR (Epidermal Growth Factor Receptor) protein. The drug provided an extremely potent response, but only in about 10% of the infected individuals.^{2,3} The ineffectiveness of single medicine paradigm necessitated the discovery of new paradigms where the drug therapy can block more than one target.

Among the different strategies developed to address the above issues, combination therapy was considered as a popular alternative in which a cocktail of drugs is co-administered in the form of two or more individual tablets to treat unresponsive patients.⁴ However, the advantage of combining therapeutic mechanisms of different drugs through this approach is compromised by patient compliance.^{5,6} The multi-component drug approach, involving co-formulation of two or more drugs in a single tablet, makes dosing regimens simpler, improves patient compliance,^{7,8} and even obviates the risk of drug-drug interactions present in combination therapy. This strategy has enhanced research and development (R & D), as evident by the launch of several multi-component drugs, including Caduet⁹ (atorvastatin + amlodipine) and Vytorin¹⁰ (simvastatin + ezetimibe) for the treatment of cardiovascular disease, Coartem (artemether + lumefantrine) and Artekin (dihydroartemisinin +

piperazine) for the treatment of malaria,¹¹ and Atripla¹² (efavirenz + tenofovir/emtricitabine) against AIDS. However, highly complex pharmacokinetic (PK)/pharmacodynamic (PD) relationships in multi-component drugs due to the differences in relative rates of metabolism of drugs in different patients led to unpredictable variability. With the failure of these strategies, the development of new drug molecules that aim to modulate multiple targets simultaneously (polypharmacology) along with enhanced efficacy, improved safety, synthetic selectivity and economic accessibility represents a big challenge for the pharmaceutical sector.

Nowadays, molecular hybridization has successfully emerged as a promising tool for medicinal chemists and the drug design process, in which two or more different pharmacophoric units are covalently linked into a single hybrid molecule with superior affinity and efficacy as compared to the parent drugs.^{13–16} Molecular hybridization is beneficial as different targets are activated by a single molecule and is particularly interesting where treatment is limited to a few commercial drugs or in cases where the discovered bioactive compounds present high toxicity or pharmacokinetic and pharmacodynamic limitations.^{17,18} Hybrid molecules can be designed in different ways as follows:

- *Metabolically stable hybrids*, in which the desired molecules are tethered *via* linkers or spacers, which may or may not be stable under *in vivo* conditions. These conjugates interact preferably at more than one target with effects mostly additive or synergistic for a particular disease.

- *Cleavable molecular hybrids*, which get metabolized *in vivo* with the release of parent drug molecules and have different targets and mechanisms of action.

- Molecular hybrids can also be prepared *via* either fusing the drug molecules without introducing a linker or by merging the individual molecules by taking advantage of commonalities in their structure.

Today, much of the world's population is affected by infectious diseases that remain instrumental in debilitating poverty. According to the latest statistics published in 2012, 8.7 million people died worldwide in 2008 due to infectious diseases.¹⁹ People who lack food, shelter, security and social protection are more vulnerable to infectious diseases since they often lack the most basic measures of prevention and care. Pathogenic microorganisms, including bacteria, viruses, parasites or fungi, are the major cause of infectious disease and can spread directly or indirectly, from person to person. Among infectious diseases, malaria, tuberculosis (TB) and HIV/AIDS are high on the global agenda while other less important infections include Chagas disease, human African trypanosomiasis, trichomoniasis and leishmaniasis.²⁰

Malaria is considered to be one of the most dangerous parasitic diseases because of its high morbidity and mortality, as well as its socio-economic impacts on the malaria-endemic region. It remains a chief cause of illness and death in tropical and subtropical countries, including Africa, Asia and South America. Approximately 90 percent cases of malaria are found in sub-Saharan Africa. According to the World Health Organization (WHO) report 2012, 3.4 billion people in 103 countries



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and extensively worked on molecular hybridization protocols for the preparation of molecular conjugates intended for HIV-malaria co-infections. At present, he is working on a research fellowship at the University of Umeå, Sweden. His research interests include the development of diverse synthetic protocols for the synthesis of novel molecular frameworks targeting tropical infections. He has also been engaged in the utilization of β -lactam synthon protocols for the synthesis of functionally decorated and biologically relevant heterocycles with medicinal potential.

are at risk of infection with 207 million malaria cases and 627 000 deaths, with the majority of victims being pregnant women or children less than the age of five years.²¹ The disease is caused by genus *Plasmodium* and, among 200 *Plasmodium* species, only five infect humans, viz. *Plasmodium falciparum*, *P. vivax*, *P. ovale*, *P. malariae* and *P. knowlesi*, with *P. falciparum* being the most virulent. Historically, the safe and cheap drug chloroquine (CQ) was extensively used for the treatment of malaria due to its excellent clinical efficacy, restricted host toxicity and simple cost-effective synthesis. CQ is believed to target ferriprotoporphyrin IX (FPIX) heme inside the digestive vacuole (DV) of the parasite. FPIX, being toxic to the parasite in its free state, is sequestered as a non-toxic crystalline hemozoin, also called "malarial pigment." CQ forms a complex with FPIX, resulting in the accumulation of toxic hemozoin (Fe(III)PPIX) within the digestive vacuole and instigating the death of the parasite.^{22–29} However, the onset of resistant strains to the available antimalarial drugs, including CQ, is responsible for the global rise of malaria and offers a strong impetus for the development of new antimalarial drugs.^{30,31} The development of resistance of *P. falciparum* to CQ is mainly attributed to the mutations in the *P. falciparum* chloroquine resistant transporter gene (PfCRT), a protein involved in the efflux of drug and proton equilibrium across the membrane of the digestive vacuole, resulting in poor accumulation of CQ in the acidic food vacuole of the parasite.^{32,33} Currently, the WHO recommends artemisinin based combination therapy (ACT) comprising artemisinin and its semi-synthetic derivatives in combination with existing drugs for the treatment of malaria. This drug regimen is considered successful against both CQ-sensitive as well as CQ-resistant strains of malaria.^{34–36} However, recent reports of the development of clinical resistance to ACT in Southeast Asia threaten this combination therapy.³⁷

4-Aminoquinoline hybridization is now considered an attractive and viable strategy for preventing and delaying the emergence of drug resistance along with the improvement in efficacy.^{38–42} The success of the quinoline-hybridization strategy was exemplified by several potential antimalarials, such as trioxaferoquinines,⁴³ trioxaquinines,⁴⁴ artemisinin–quinine hybrid,⁴⁵ 4-aminoquinoline based tetraoxanes,⁴⁶ clotrimazole-based-4-aminoquinoline⁴⁷ and isatin–4-aminoquinoline hybrids.⁴⁸ The present review article encompasses the recent developments on the utility of 4-aminoquinoline-hybridization towards the development of novel antimalarials. A special focus has been given to the structure–activity relationship, IC₅₀ values against CQ-sensitive and resistant strains, *in vivo* evaluation data and cytotoxic studies of the promising candidates that have emerged from this strategy.

2. 4-Aminoquinoline-based antimalarial conjugates

2.1 4-Aminoquinoline–chalcone conjugates

Chalcones and dienones are structurally linked compounds that have been revealed to exhibit notable *in vitro* and *in vivo* antimalarial activity^{49–52} by acting as inhibitors of either

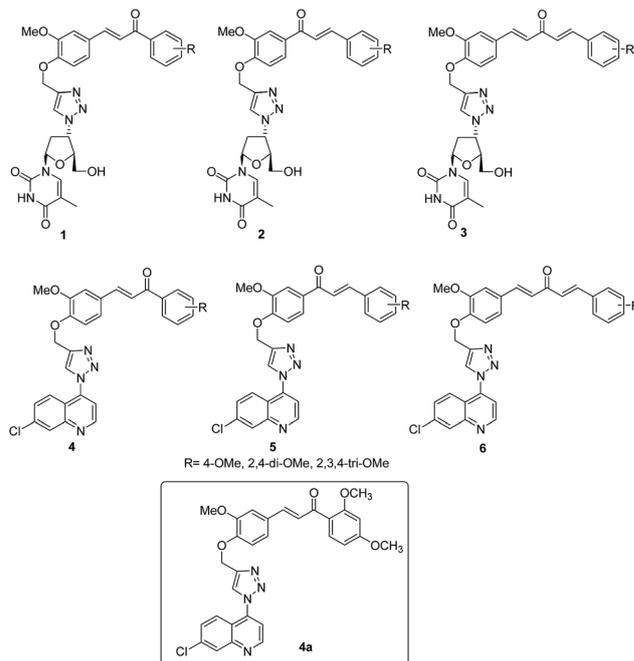
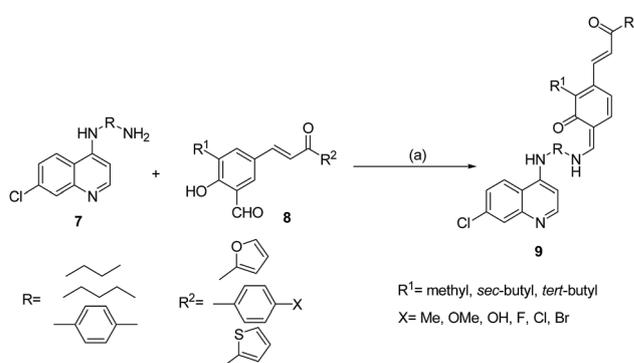
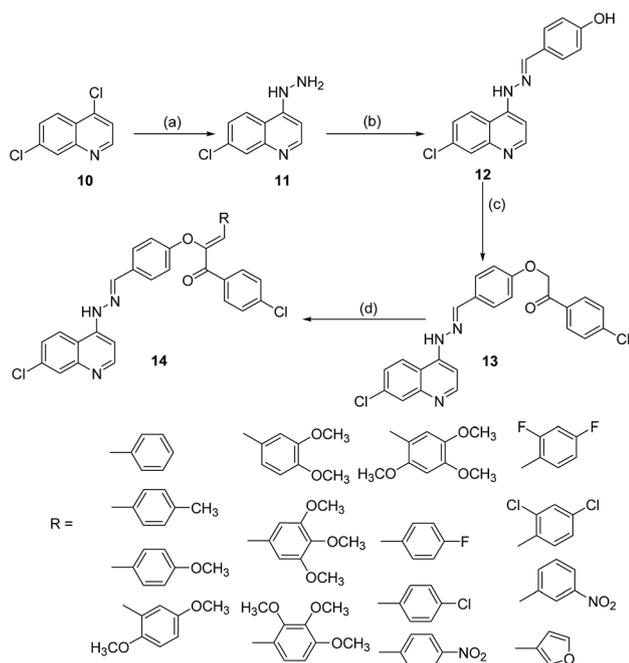


Fig. 1 General structures for triazole-linked chalcone and dienone hybrid compounds comprising AZT and aminoquinoline with the most active hybrid being 4a.

plasmodial aspartate proteases,⁵³ cysteine proteases⁵⁴ or permeability pathways initiated into erythrocyte cell membranes by the malaria parasite.⁵⁵ Chibale and co-workers applied the molecular hybridization strategy *via* Cu(I)-catalyzed cycloaddition reaction of terminal alkynes and azides for the synthesis of series of 1*H*-1,2,3-triazole linked chalcone and dienone conjugates containing aminoquinoline and nucleoside templates (Fig. 1).⁵⁶ The synthesized hybrids were screened for their antimalarial activity against the CQ-sensitive (D10) and CQ-resistant (Dd2 and W2) strains of *P. falciparum*. Notably, the azidothymidine (AZT) conjugates with both chalcones and dienones did not show any improvement in the antimalarial activity over their acetylenic precursors while retention of activity was observed in most cases. The quinoline-hybridization approach led to the identification of highly



Scheme 1 Reagents and conditions: (a) ethanol, rt, 10 min.



Scheme 2 Reagents and conditions: (a) hydrazine hydrate, EtOH, reflux, 8 h; (b) 4-hydroxybenzaldehyde, EtOH, reflux, 2–3 h; (c) 4-chlorophenacyl bromide, K_2CO_3 , acetonitrile, rt, 2 h; (d) different substituted aldehydes, 10% methanolic KOH, rt.

potent hybrids, with the most active conjugate **4a** having sub-micromolar IC_{50} values of 0.04, 0.07 and 0.09 μM against tested D10, Dd2 and W2 strains of *P. falciparum*, respectively. Cytotoxicity against the Chinese Hamster Ovarian cell line was determined and the hybrid **4a** proved to be non-cytotoxic even at the highest concentration tested (100 μM).

Sashidhara and co-workers utilized an uncommon approach towards the synthesis of keto-enamine tethered chalcone–4-aminoquinoline conjugates (**9**).⁵⁷ The synthesized conjugates were evaluated for their antimalarial potential against a CQ-sensitive (3D7) strain of *P. falciparum* (Scheme 1). The promising compounds from *in vitro* assay were further screened for their antimalarial efficacy against *P. yoelii* (CQ-resistant N-67 strain) in Swiss mice. Two of the synthesized conjugates displayed suppression of 99.9 percent parasitemia on day 4. Mechanistically, the test compounds exhibited an antimalarial mode of action similar to that of CQ, as confirmed by inhibition of β -hematin formation studies.

The above work was further extended towards the synthesis of 4-aminoquinoline–chalcone conjugates by following the sequence of synthetic steps shown in Scheme 2 along with their antimalarial evaluation.⁵⁸ Most of the synthesized conjugates showed improved antimalarial profile against the Q-resistant K1 strain when compared to CQ. Structure activity relationship (SAR) studies demonstrated the dependence of the activity profile on the substitution pattern as well as the number of substituents present on the phenyl ring. The presence of electron releasing groups ($R = -CH_3, -OCH_3$) has been shown to improve the activity profiles while replacing them with electron

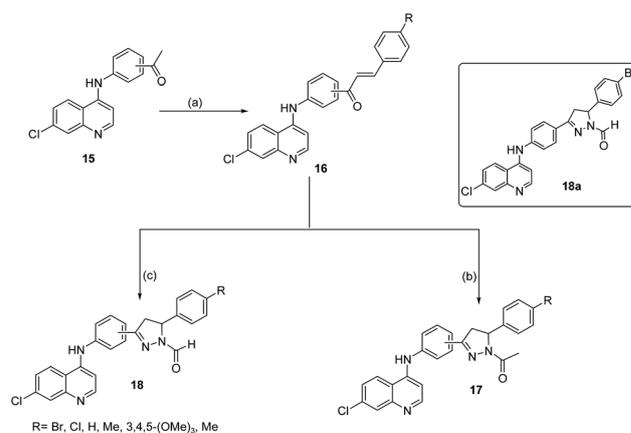
withdrawing groups ($R = -Cl, -NO_2$) reduced the antimalarial activity, with the exception of fluorine. β -Hematin studies were also carried out for the synthesized conjugates with the three compounds being most active in inhibition of β -hematin formation with IC_{50} values of 3.46, 3.52 and 3.74 $\mu g\ ml^{-1}$.

Insuasty *et al.* utilized 7-chloroquinoline–amino-chalcone hybrids (**16**) for the preparation of a series of *N*-acetyl and *N*-formyl-pyrazoline derivatives (**17** and **18**) in acceptable to good yields *via* a cyclo-condensation reaction using hydrazine hydrate under acidic conditions (Scheme 3). The synthesized conjugates were bio-evaluated for their antimalarial and anti-cancer profiles.⁵⁹ The anticancer profiles against 60 cell lines revealed that GI_{50} values for most of these conjugates range from 0.13 to 0.99 μM . The antimalarial activity performed against the NF54 strain of *P. falciparum* revealed that one of the *N*-formyl-pyrazoline derivatives, **18a**, exhibited 50.8% inhibition.

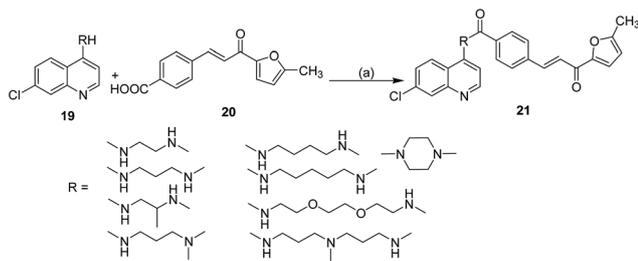
A series of 4-aminoquinolinyl-chalcone amides (**21**) were synthesized by N'Da through condensation of carboxylic acid-functionalized chalcone with aminoquinolines (Scheme 4) along with their screening against CQ-sensitive (3D7) and CQ-resistant (W2) strains of *P. falciparum*. Cytotoxicity against the WI-38 cell line of normal human fetal lung fibroblast was also evaluated.⁶⁰ The IC_{50} values of the synthesized conjugates ranged between 0.04–0.5 μM and 0.07–1.8 μM against 3D7 and W2, respectively. SAR studies revealed the increased antimalarial activity of the amides with the increase in lipophilicity and alkyl chain length. Moderate to high toxicity towards mammalian cells was observed for these conjugates. The most active compound featuring 1,6-diaminohexane as the linker was two-fold more potent than CQ against the 3D7 and W2 strains despite its predicted high lipophilicity, low solubility and poor absorption properties.

2.2 4-Aminoquinoline–pyrimidine conjugates

Rawat and co-workers reported the preparation of a library of 4-aminoquinoline–pyrimidine conjugates in an attempt to



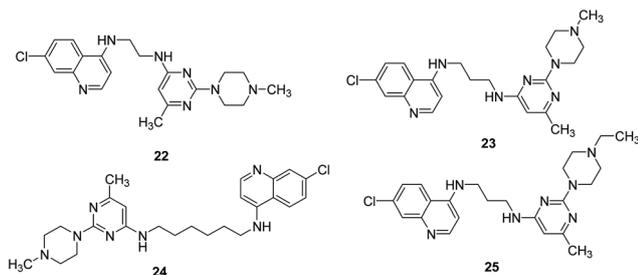
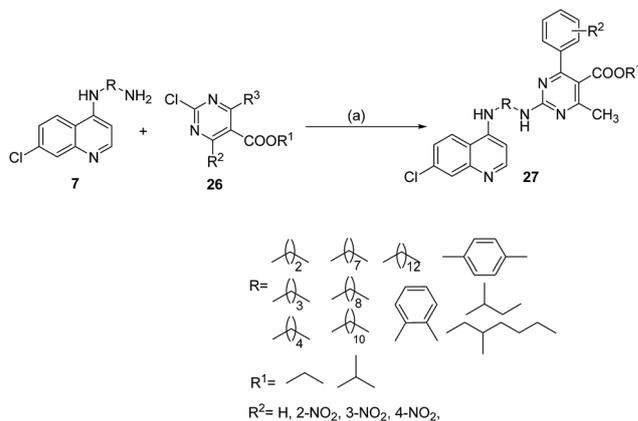
Scheme 3 Reagents and conditions: (a) KOH, MeOH, rt; (b) hydrazine hydrate, acetic acid, reflux; (c) hydrazine hydrate, formic acid, DMF, reflux.



Scheme 4 Reagents and conditions: (a) CDI, DCM : DMF, rt, 24 h.

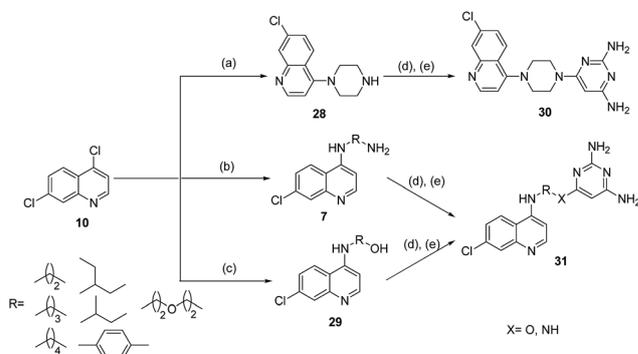
search for more active molecules with effectiveness against both CQ-sensitive and CQ-resistant strains of *P. falciparum*.⁶¹ Among the synthesized compounds, eleven conjugates displayed enhanced antimalarial activity compared to CQ against both D6 and W2 strains, while four conjugates (**22**, **23**, **24**, and **25**, Fig. 2) showed better activity against both CQ-sensitive as well as CQ-resistant (D6 and W2) strains of *P. falciparum* in comparison to pyrimethamine. Most of the conjugates were found to be non-cytotoxic up to a concentration of 60 μM , while others showed mild toxicities. Most of the conjugates exhibited a high selective index; compounds **22** and **25**, with IC_{50} values of 0.005 and 0.006 μM , respectively, against the D6 strain and 0.03 and 0.06 μM , respectively, against the W2 strain of *P. falciparum*, were selected for *in vivo* studies, and showed outstanding activity in a mouse model of *P. berghei* without any toxicity.

In a recent communication, Singh and co-workers showed the synthesis of molecular conjugates **27** based on 7-chloro-4-aminoquinoline (**7**) and 2-aminopyrimidine (**26**) motifs, as shown in Scheme 5.⁶² The rationale behind using 2-aminopyrimidine was because of its well established antimalarial potential, as evidenced by the structurally related drug pyrimethamine. The antiplasmodial profiles of the synthesized conjugates were in the nanomolar range, with the most potent hybrid exhibiting an IC_{50} value of 3.6 nM, 56-fold less compared to CQ against the CQ-resistant K1 strain. Almost all of the synthesized compounds were cytotoxic and the binding studies with DNA implied their strong affinity for target parasite type AT rich pUC18 DNA. The active conjugates also showed good inhibitory activity for β -haematin formation, suggestive of the fact that the observed antiplasmodial potential of the conjugates in the present case is because of their ability to act on multiple targets.

Fig. 2 Most potent 4-aminoquinoline-pyrimidine conjugates **22**, **23**, **24** and **25**.Scheme 5 Reagents and conditions: (a) K_2CO_3 , THF, 48 h, rt.

A range of 7-chloroquinoline-pyrimidine hybrids, *viz.* **30** and **31**, were synthesized by N'Da and co-workers *via* aromatic nucleophilic substitution reaction of 4-aminoquinolines with 2,6-diamino-4-chloropyrimidine and their antiplasmodial potential evaluated. The conjugates were evaluated alongside CQ, pyrimethamine (PM) and their fixed combinations *viz.* CQ/PM (1 : 1) and CQ/PM (1 : 4) against the CQ-susceptible D10 and -resistant Dd2 strain of *P. falciparum* (Scheme 6).⁶³ The cytotoxic profiles of the synthesized scaffolds were also evaluated against the Chinese Hamster Ovarian (CHO) cell line. All the synthesized hybrids showed activity against both D10 and Dd2 strains with good selective index. The most potent compound of the series **30** with piperazine as the linker showed comparable activity to that of PM and CQ against the D10 strain ($\text{IC}_{50} = 0.07 \mu\text{M}$) while a three-fold better potency than that for CQ against Dd2 strain ($\text{IC}_{50} = 0.157 \mu\text{M}$) was observed.

Further, a library of piperazine tethered 4-aminoquinoline-pyrimidine conjugates was prepared and screened for their antimalarial profiles over CQ-sensitive (D6) and CQ-resistant (W2) strains of *P. falciparum* while cytotoxicity was evaluated against the mammalian Vero cell line.⁶⁴ Nine conjugates were shown to have superior antimalarial potency against both the

Scheme 6 Reagents and conditions: (a) piperazine, DMF, 80–135 $^\circ\text{C}$, 5 h; (b) diaminoalkane/aryldiamine, neat or DMF, 80–150 $^\circ\text{C}$, 24 h; (c) aminoalcohol, neat, 120 $^\circ\text{C}$, 24 h; (d) NaH, DMF, rt, 1 h; (e) 2,6-diamino-4-chloropyrimidine, DMF, 135 $^\circ\text{C}$, 16–24 h.

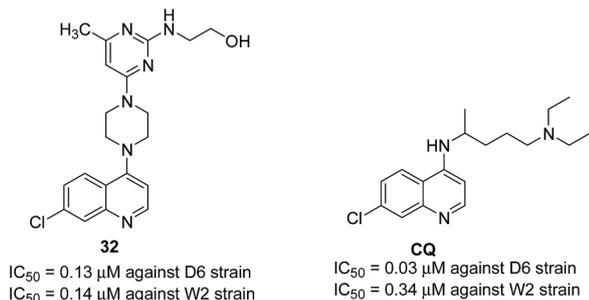


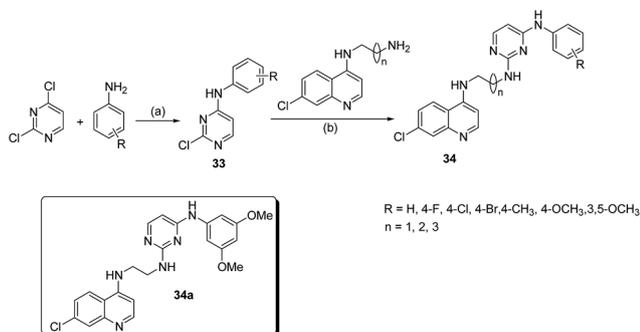
Fig. 3 Most potent piperazine tethered 4-aminoquinoline–pyrimidine conjugate **32** and **CQ**.

strains with IC_{50} values ranging from 0.13 to 0.14 μM . Out of these, compound **32** (Fig. 3) proved to be the most potent among the synthesized conjugates and its antimalarial activity was found to be 2.5 fold more than that of the standard drug **CQ**. All the synthesized conjugates were non-cytotoxic against the Vero cell line.

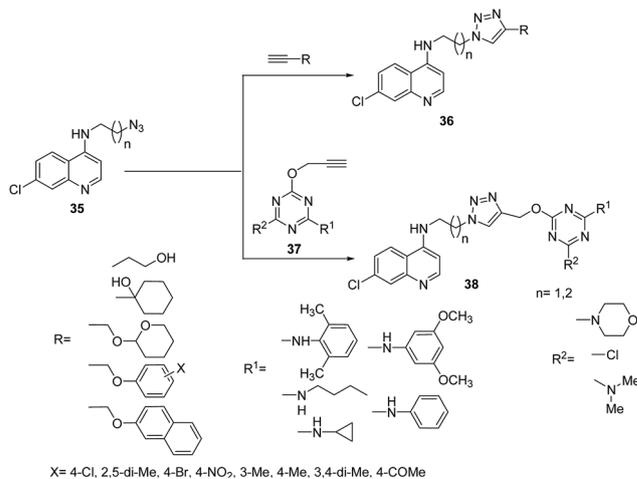
The above work was further extended for the preparation of 4-aminoquinoline–pyrimidine hybrids **34**, as shown in Scheme 7. All the synthesized compounds were screened for their antiplasmodial activity against D6 and W2 strains of *P. falciparum*, depicting activity in the nano-molar range.⁶⁵ Numerous compounds were found to be nontoxic to the mammalian cell lines and showed promising *in vitro* antimalarial activity over both **CQ**-sensitive and -resistant strains with high values of selective index. The most potent conjugate **34a** with an IC_{50} value of 0.033 and 0.058 μM against D6 and W2 strains of *P. falciparum*, respectively, was chosen for *in vivo* studies and exhibited significant suppression of parasitemia. The heme binding studies confirmed heme as one of the possible targets of these hybrids with heme forming a stable 1 : 1 complex with these conjugates. Potent compounds from *in vitro* antimalarial activity data were selected for molecular docking simulations and showed good interaction with the binding sites of PfDHFR.

2.3 4-Aminoquinoline–triazine conjugates

Rawat *et al.* described the synthesis of a library of 4-aminoquinoline-1,2,3-triazoles (**36**) and 4-aminoquinoline-



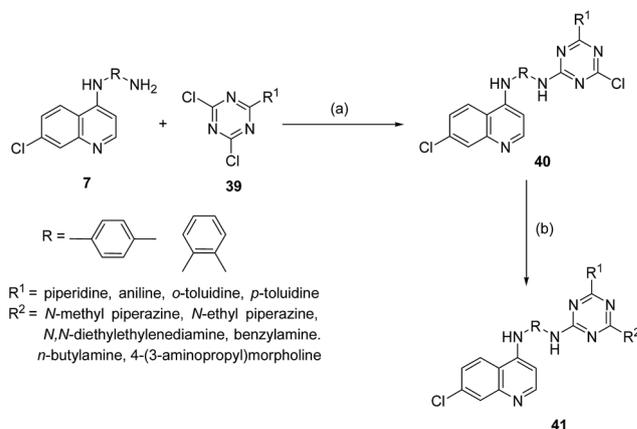
Scheme 7 Reagents and conditions: (a) TEA, EtOH, rt, overnight; (b) K_2CO_3 , NMP, 140–150 °C, 10–12 h.



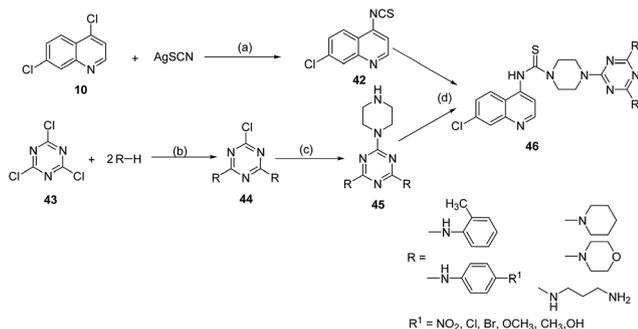
Scheme 8 Reagents and conditions: (a) $CuSO_4 \cdot 5H_2O$, sodium ascorbate, *t*-BuOH : H₂O (1 : 1), 40 °C, 3 h.

1,2,3-triazole–1,3,5-triazine conjugates (**38**) by utilizing Huisgen-1,3-dipolar cycloaddition reaction of 4-azido-7-chloroquinoline with variedly substituted terminal alkynes (Scheme 8).⁶⁶ It has been hypothesized that amalgamation of a basic nucleus *viz.* triazine with 4-aminoquinoline would enhance the accumulation of the conjugate in the digestive food vacuole, targeting *P. falciparum* dihydrofolate reductase (PfDHFR). However, the antimalarial profile of the synthesized conjugates against **CQ**-sensitive D6 and **CQ**-resistant W2 strains revealed that none of the hybrids exhibited better activity than that of **CQ**.

The above study was further extended by Chauhan and co-workers in the synthesis of 4-anilinoquinoline-triazine derivatives (**40** and **41**) using readily available and cheap starting materials (Scheme 9). The compounds were assessed for their antiplasmodial efficacy against the **CQ**-sensitive 3D7 strain of *P. falciparum* as well as for their cytotoxicity towards the mammalian Vero cell line.⁶⁷ The most promising and non-cytotoxic compounds from the preliminary studies were chosen for *in vivo* screening in Swiss mice infected with the



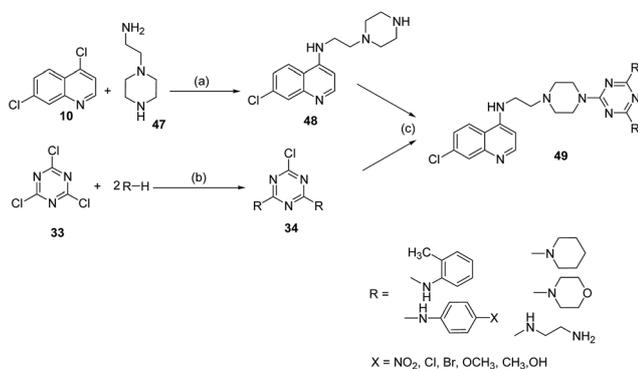
Scheme 9 Reagents and conditions: (a) mono-substituted triazines, THF, reflux, 8 h; (b) various amines, THF, reflux, 5 h.



Scheme 10 Reagents and conditions: (a) reflux with stirring for 18 h at 90–120 °C; (b) dry acetone, 0–5 °C, 1 h, then 40–45 °C for a further 3 h; (c) piperazine, 1,4-dioxane, 120–130 °C, 5–6 h; (d) dry acetone, 40–45 °C, 18 h.

CQ-resistant N-64 strain of *P. yoelli*. These studies resulted in the identification of two orally active compounds with piperidine functionality. The results obtained are suggestive of the fact that the antimalarial activity in these conjugates can be attributed to the metabolic stability of piperidine functionality. The selected conjugates were further investigated for their ability to inhibit β -hematin formation. All the tested hybrids exhibited better inhibitory activities of β -haematin formation (IC_{50} ranging from 2.65 to 3.16 μ M) than CQ (IC_{50} = 3.65 μ M), which revealed that, apart from targeting heme, there might be the possibility of other mechanisms of action.

The use of triazine functionality for the synthesis of molecular conjugates with antimalarial potential was further elaborated by Bhat and co-workers in the synthesis of variedly functionalized 1,2,3-triazine-4-aminoquinoline conjugates (46) attached *via* thiourea functionality as the linker (Scheme 10). The determination of their antimalarial efficacy was done against the CQ-sensitive (3D7) and CQ-resistant (RKL-2) strains of *P. falciparum*.⁶⁸ The observed SAR revealed a preference for electron withdrawing substituents, *viz.* bromo and hydroxyl groups, while the presence of electron donating substituents, including methyl and methoxy, adversely affected the activity profiles. The observed activities were further substantiated by



Scheme 11 Reagents and conditions: (a) reflux 1 h at 80 °C followed by 6–8 h at 120–130 °C; (b) 1,4-dioxane 0–5 °C, 1 h, 40–45 °C, 3 h; (c) 1,4-dioxane, 120–130 °C, 6–7 h.

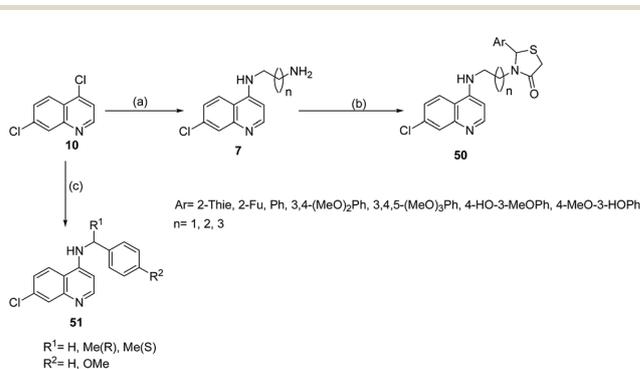
docking simulations performed by using the ligand fit module within Discovery Studio 2.5 on wild and quadruple mutant strains of *P. falciparum* dihydrofolate reductase thymidylate synthase (pf-DHFR-TS).

Further extension of the above work resulted in the synthesis of 4-aminoquinoline-1,3,5-triazine derivatives (49) connected through non-ionizable covalent linkers (Scheme 11).⁶⁹ The antiplasmodial profile again showed a similar SAR to that observed earlier with a marked preference for electron withdrawing substituents for good activity. Further, addition of basic moieties, such as morpholine, piperidine and 1,3-diaminopropane, in the synthesized conjugates was shown to enhance the antimalarial activity. The evaluation results were correlated using docking studies carried out on wild and quadruple mutants pf-DHFR-TS. In the wild type strain, most of the hybrids revealed a hydrophobic interaction between the phenyl ring of conjugates with the Phe58 residue as well as a σ - π interaction between the 1,3,5-triazine ring and the Leu46 residue. Similarly, a hydrophobic interaction was observed between the phenyl ring and Phe58 in addition to H-bond formation between the 1,3,5-triazine ring and Ser111 in the quadruple mutant of pf-DHFR-TS. Further, it was observed that the quinoline moiety was engaged in the formation of a $\pm\pi$ interaction with Arg59.

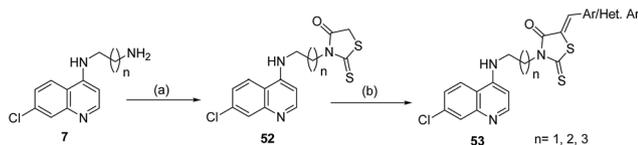
2.4 4-Aminoquinoline-thiazolidine conjugates

An alternative pathway for obtaining new and promising anti-malarial compounds was disclosed by Kouznetsov and co-workers. A library of 21 heterocyclic scaffolds consisting of either *N*-(aminoalkyl)thiazolidin-4-one-4-aminoquinoline conjugates (50) or substituted *N*-benzylamino-7-chloroquinolines (51) was prepared by following the sequence of steps depicted in Scheme 12.⁷⁰ The antimalarial evaluation against 3D7 and Dd2 strains showed that four of the *N*-benzylamino-7-chloroquinoline derivatives (51) are up to 3-fold more active compared to the standard drug, CQ. Non-specific cytotoxicity assay on J774 murine macrophages and HepG2 cells (human hepatocellular carcinoma cell line) proved their high selectivity index and hence the potential for *in vivo* evaluation.

Chauhan and co-workers recently explored the synthesis of 4-aminoquinoline-rhodanine hybrids along with their *in vitro*



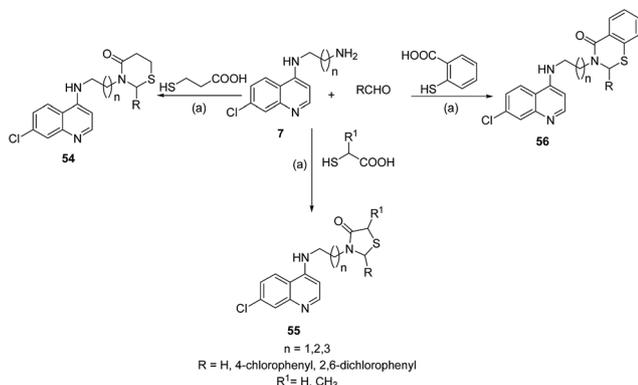
Scheme 12 Reagents and conditions: (a) diaminoalkane, 80 °C for 1 h, 140–150 °C for 6–7 h; (b) ArCHO, HSCH₂COOH, PhMe, 4–5 h reflux; (c) substituted *N*-benzylamine, K₂CO₃, DMF, 140 °C, 10 h.



Scheme 13 Reagents and conditions: (a) CS_2 , $\text{BrCH}_2\text{COOC}_2\text{H}_5$, acetonitrile, rt, 3–5 h, (b) acetic acid, ammonium acetate, aromatic/heteroaromatic aldehydes, 90°C , 4–6 h.

antimalarial efficacy against K1 and 3D7 strains of *P. falciparum*, as well as their cytotoxicity against the Vero cell line (Scheme 13).⁷¹ Although none of the hybrids was more potent than CQ against the 3D7 strain, four of the tested conjugates showed antimalarial activity comparable to CQ with IC_{50} values in the range of 13.2–45.5 nM against K1 strain along with high selectivity. Furthermore, some of the hybrids were also tested for their anti-mycobacterial potential against the H_{37}Rv strain of *M. tuberculosis* with three of the potent compounds exhibiting minimum inhibitory concentration (MIC) values of 6.25 μM . In addition, inhibition of β -hematin formation studies disclosed that the test conjugates were more active than CQ in the inhibition of hemozoin formation, clearly validating their mechanism of action.

A series of differently substituted 4-aminoquinolines having thiazolidin-4-ones (54), [1,3]thiazinan-4-ones (55) or 2,3-dihydrobenzo[*e*][1,3]thiazin-4-ones (56) at the terminal amino functionality was synthesized *via* a three-component reaction between 4-aminoquinolines, aryl aldehydes and appropriate mercapto acids (Scheme 14).⁷² Among the compounds tested against the NF-54 strain of *P. falciparum*, nine showed IC_{50} values ranging between 0.013–0.98 μM . The promising compounds from *in vitro* assay were further selected for *in vivo* activity testing in Swiss mice using N-67 strain of *P. yoelli*. The most potent compound from *in vivo* analysis suppressed parasitemia by 81 percent on day 4 compared to 100 percent suppression exhibited by CQ. SAR studies showed that the lateral side chain modification of 4-aminoquinoline in these conjugates was well tolerated with the best results obtained with a two- or three-carbon chain as the linker. The biochemical studies confirmed their association with hematin and hence confirmed the mechanism of action of the test compounds.



Scheme 14 Reagents and conditions: (a) DCC, THF, rt or PhMe, reflux.

2.5 4-Aminoquinoline–clotrimazole conjugates

With the aim of developing new antimalarials, in a recent communication Gemma and co-workers identified a new polyaromatic antimalarial pharmacophore based on a clotrimazole scaffold.⁷³ Clotrimazole is a popular antimycotic drug with weak antimalarial activity ($\text{IC}_{50} = 0.55 \mu\text{M}$) against the W2 strain of *P. falciparum*. However, its peculiar structural features, *viz.* (i) the presence of an imidazole nucleus, identified to mediate electron transfer reaction and (ii) the triphenylmethyl system known to stabilize radical intermediate, makes it an ideal candidate to interact with the haemoglobin-derived ferroprotoporphyrin ($\text{Fe(II)}\text{-FP}$) complex in the food vacuole of the parasite. Consequently, a number of polyaromatic pharmacophores structurally related to clotrimazole have been synthesized with an extension towards clotrimazole–4-aminoquinoline conjugates. The antimalarial evaluation of the synthesized scaffolds showed dependence upon some key structural features, *viz.* (i) the protonatable side chain, (ii) the imidazole ring, and (iii) the aryl/heteroaryl system. The most promising compounds (57, 58 and 59 in Fig. 4) were further evaluated to assess their *in vivo* efficacy against *P. chabaudi* in CDI mice after oral administration in a 4 day suppression test. The results showed that compounds 57 and 58 have low propensity to produce rapid resistance along with *in vivo* activity against plasmodia and oral bioavailability.

The above protocol was further extended towards the synthesis of a series of antimalarial hybrids by combining the 4-aminoquinoline unit with that of clotrimazole, as depicted in Fig. 5.⁷⁴ The antimalarial studies were carried out *in vitro* against CQ-sensitive and -resistant strains as well as *in vivo* in a rodent malaria model. Compounds 63 and 64 displayed strong antimalarial activity against *P. berghei* after oral administration. The compounds have been shown to interfere with the process of heme detoxification, as confirmed by BHIA assay. Cytotoxicity, genotoxic potential and effects on cardiac function of lead compounds were also assessed with the compound 63 emerging as a good hit because of its promising antimalarial potency and an optimal half-life in mice.

2.6 4-Aminoquinoline–azithromycin conjugates

Azithromycin, a semi-synthetic macrolide antibiotic, was recently discovered as a slow acting antimalarial^{75,76} that yields its activity *via* inhibiting protein synthesis on prokaryote-like ribosomes in *P. organelle*.⁷⁷ Currently, it is in clinical trials and has been used as a combination partner with different

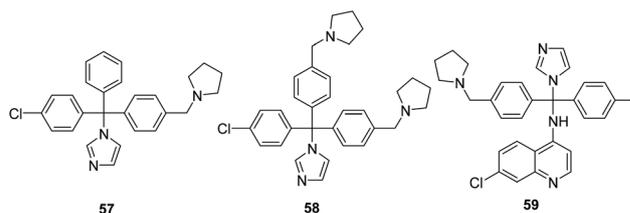


Fig. 4 Most potent compounds 57, 58 and 59.

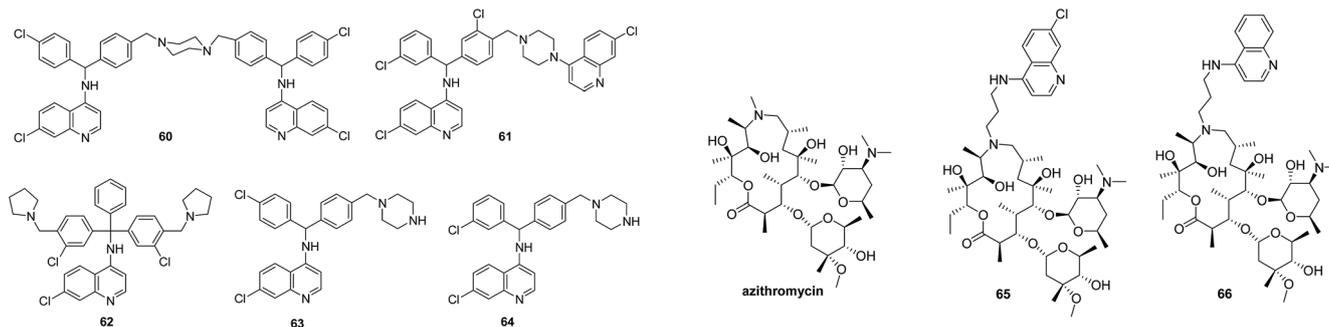


Fig. 5 Molecular conjugates **60**, **61** and **62** of 4-aminoquinoline and clotrimazole along with their potent precursors **63** and **64**.

antimalarials. Perić and collaborators employed a molecular hybridization strategy for the synthesis of azithromycin–quinoline antimalarial conjugates.⁷⁸ Forty two new azithromycin analogues and azithromycin–quinoline conjugates with amide and amine functionalities were synthesized using simple and economical chemical procedures. These scaffolds displayed 100-fold improvement in *in vitro* potency with high selectivity, pharmacokinetics (PK) and *in vivo* efficacy over azithromycin against the *P. falciparum* TM91C235 strain, a multidrug resistant clone from Southeast Asia. The screening of the promising compounds in a mouse efficacy model resulted in the identification of five scaffolds, *viz.* three amine (**65**, **66**, **67**) and two amide (**68**, **69**) derivatives, exhibiting better *in vivo* efficacy compared to azithromycin (Fig. 6). The most potent of the tested compounds **65** (Fig. 6) showed 100-fold better *in vitro* activity with excellent pharmacokinetic parameters curing mice more efficiently than azithromycin.

Perić *et al.* further extended this concept towards the development of the azalide class of antimalarials *via* synthesis of 2'-*o*-substituted-9-deoxy-9a-methyl-9a-aza-9a-homoerythromycin A derivatives covalently linked to varied substituted quinolines at position 2', as depicted in Fig. 7.⁷⁹ Antimalarial profiles of the test conjugates against the CQ-sensitive 3D7 strain showed 100-fold improvement over azithromycin while 48-fold enhancement was observed against the CQ-resistant W2 strain of *P. falciparum*. These results have facilitated these macrolides for the preclinical development of malaria-specific agents.

2.7 4-Aminoquinoline– β -lactam conjugates

A series of 1*H*-1,2,3-triazole-linked 7-chloroquinoline– β -lactam conjugates (**72** and **73**) was synthesized by Kumar and co-workers utilizing click chemistry, as shown in Fig. 8. The antimalarial potency of the synthesized conjugates against a W2-resistant strain of *P. falciparum* was found to be dependent on the substituent present at the *N*-1 position of the β -lactam ring and the presence of bis-triazole at C-3 position. The observed antimalarial efficacy was further validated by docking studies through inhibition of *P. falciparum* dihydrofolate reductase (PfDHFR).⁸⁰

On the basis of these preliminary results, Kumar *et al.* further extended their work on 4-aminoquinoline hybridization by synthesizing 4-aminoquinoline– β -lactam conjugates linked

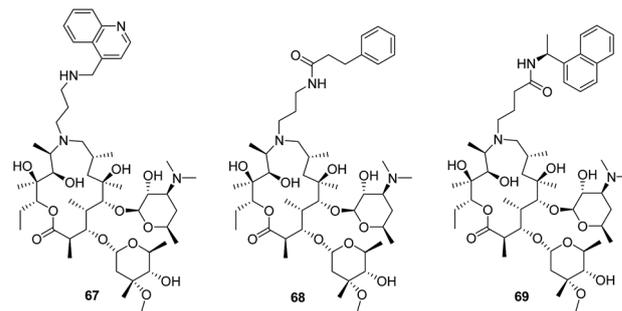


Fig. 6 Structure of azithromycin and potent azalide hybrids used for *in vivo* study in the mouse model.

via a diverse range of linkers, *viz.* amide⁸¹ (**75**, Fig. 8) and non-ionizable covalent bonds (**74**, Fig. 8),⁸¹ urea⁸² (**76**, Fig. 8) and oxalamide⁸² (**77**, Fig. 8) functionalities along with well-modulated alkyl chain lengths. All the synthesized conjugates were screened for their antimalarial potential against a W2 resistant strain of *P. falciparum*. Antiplasmodial data revealed that the activity depends on the nature of the linker, the alkyl chain length and the substituent present at the *N*-1 position of the β -lactam ring. On comparing the potencies among the amide and alkyl chain tethered series, the conjugates linked *via* non-ionizable covalent alkyl chain linker proved to be better in inhibiting the growth of *P. falciparum* while the introduction of amide functionality adversely affected the activity profiles. The introduction of urea/oxalamide functionalities, on the other hand, resulted in improvement of the antiplasmodial profiles. The most potent and non-cytotoxic conjugate (**77a**, Fig. 8) among the 7-chloroquinoline– β -lactam series with the best

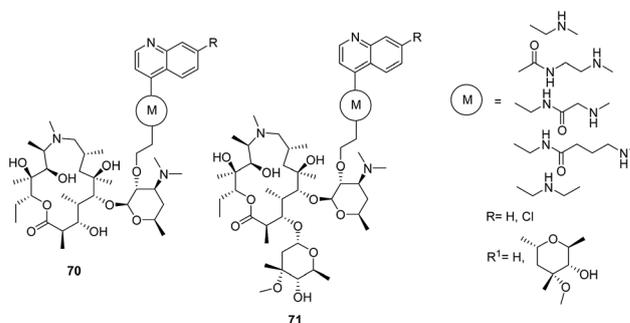


Fig. 7 General structure of hybrids of 2'-*O*-substituted-9-deoxy-9a-methyl-9a-aza-9a-homoerythromycin A and quinolone.

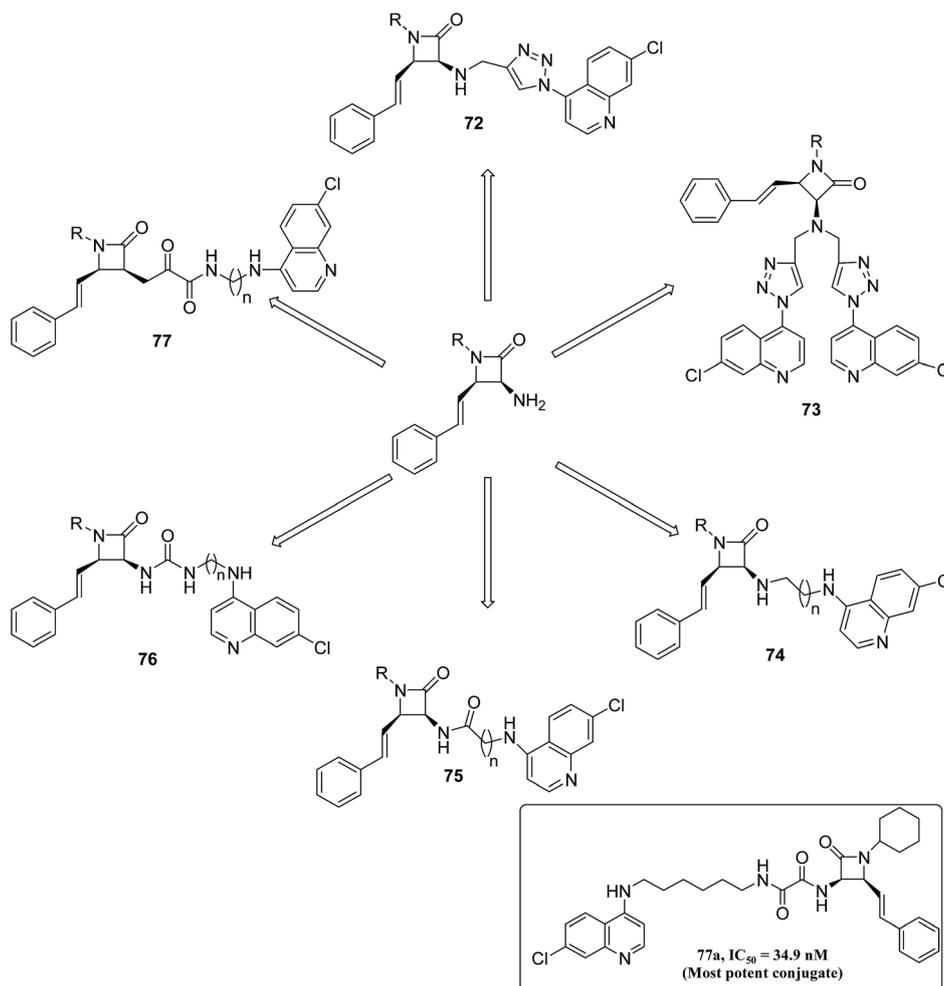


Fig. 8 β -Lactam-4-amino-quinoline conjugates.

combination of *N*-cyclohexyl substituent at *N*-1, alkyl chain length ($n = 6$) and oxalamide functionality exhibited an IC_{50} of 39.9 nM.

Further, Kumar *et al.* synthesized a library of C-3 thiourea tethered β -lactam-7-chloroquinoline conjugates (78) along with the unexpectedly formed 7-chloroquinoline-thiohydantoin derivatives (79) with the aim of searching antimalarial structure-activity relationships (Fig. 9). The synthesized thiourea-tethered 7-chloroquinoline- β -lactam conjugates 78 were found to be ineffective in inhibiting the growth of *P. falciparum*. However, the unexpected formed product *viz.* 7-chloroquinoline-thiohydantoin 79 demonstrated nanomolar antimalarial activities against the W2 strain of cultured *P. falciparum*, with the most potent and non-cytotoxic compound exhibiting an IC_{50} of 39.8 nM. β -Hematin formation studies strongly supported the mechanism of their action *via* the inhibition of β -hematin formation having IC_{50} values comparable to those of CQ.⁸³

2.8 4-Aminoquinoline-isatin conjugates

Santos and colleagues designed, synthesized and evaluated 3-methylene-substituted indolinones (Fig. 10, 80-85) as

falcipain inhibitors and antiplasmodial agents.⁸⁴ Various indolinones with a Leu-i-amyl recognition moiety were shown to display fair inhibitory activity of *P. falciparum* cysteine protease falcipain-2 and antimalarial potency against the W2 strain of *P. falciparum* in the low μ M range. Importantly, compounds lacking the recognition moiety were devoid of FP-2 inhibition and interaction of the Leu-i-amyl sequence with the S2 pocket. Further, the introduction of a 4-aminoquinoline to the C-3 position of the indolinone-2-one scaffold (85) led to a considerable enhancement in the antiplasmodial activity, suggestive of the fact that the 3-methylene-indolinone-2-ones could become forthcoming lead compounds for the development of new antimalarials.

Kumar *et al.* synthesized a library of isatin-7-chloroquinoline conjugates linked by 1*H*-1,2,3-triazole moiety along with assessment of their antiplasmodial profiles (Scheme 15). The tested compounds (88) were devoid of antiplasmodial activity, while 89, having an alkyl chain between isatin and 7-chloroquinoline moieties, displayed enhancement in antiplasmodial potency. Activity was found to depend on the C-5 substituent of the isatin ring and on the alkyl chain length between the isatin and 7-chloroquinoline moieties. The most potent conjugate,

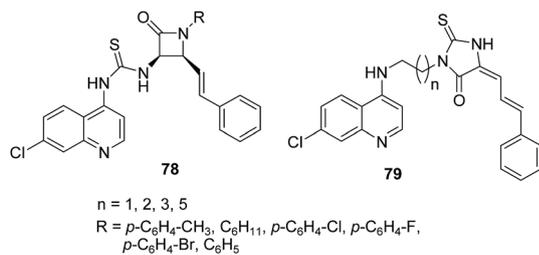


Fig. 9 General structures of β -lactam-7-chloroquinoline conjugates (78) and 7-chloroquinoline-thiohydantoin derivatives (79).

with the best combination of propyl linker ($n = 3$) and chloro-substituent at the C-5 position of isatin ring, exhibited an IC_{50} value of $1.21 \mu\text{M}$.⁸⁵

The above work was further extended towards the synthesis of triazole linked isatin-7-chloroquinoline (93) and 3-hydroxy-indole-7-chloroquinoline hybrids (94) by following the synthetic protocol shown in Fig. 11 along with their anti-plasmodial evaluation against the CQ-resistant W2 strain.⁸⁶ studies revealed the dependence of activities on the length of alkyl chain but independent of the nature of the substituent present at the C-5 position of the isatin or indole ring. Most of the conjugates were less active than the standard drug CQ; however, the best conjugate with an optimum combination of 3-hydroxy-indole ring and *n*-butyl linker exhibited an IC_{50} value of 69 nM, comparable to that of CQ. Further, Kumar and co-workers synthesized piperazine tethered 7-chloroquinoline-isatin conjugates (95 and 96, Fig. 12) by both direct nucleophilic substitution and Cu(I)Cl mediated Mannich reaction.⁸⁷ All the synthesized conjugates were assessed for their anti-plasmodial, anti-tubercular and cytotoxic evaluation. Analysis of anti-plasmodial activity data against the W2 strain showed that none of conjugates were as active as CQ, although some conjugates proved to have good anti-malarial efficacy with IC_{50} s ranging from 0.22 to $0.27 \mu\text{M}$.

Anti-tubercular evaluation against *M. tuberculosis* revealed the dependence of the activity profile on the presence of a substituent at the C-5 position of the isatin ring whereas longer alkyl chain lengths badly affected the efficacy of the compounds. Compound 95, with a fluoro substituent, was shown to have good anti-TB efficacy with six times more potency than the standard drug, Cephalexin. On the other hand, Mannich adducts (96) were found to be ineffective in inhibiting the growth of *M. tuberculosis*. The most potent compound 95a (Fig. 12), with an IC_{50} of $0.22 \mu\text{M}$ against the CQ-resistant strain of *P. falciparum* and $31.62 \mu\text{M}$ against the 3T6 cell line, exhibited a high selectivity index.

Recently, Kumar *et al.* synthesized β -amino alcohol tethered isatin-4-aminoquinoline conjugates with the aim of investigating their antimalarial structure activity relationship. SAR studies have shown a clear preference for shorter alkyl chain lengths ($n = 2, 3$) and the presence of a Cl-substituent at the C-5 position of the isatin ring for good activity. The two most potent and non-cytotoxic conjugates (97 and 98, Fig. 13) displayed antiplasmodial potency comparable to that of CQ, with IC_{50}

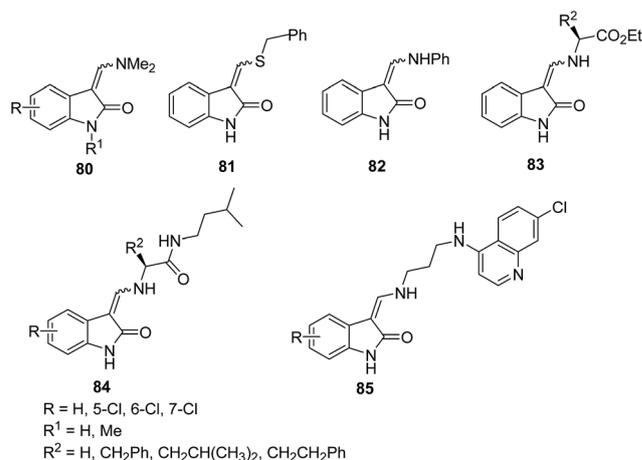


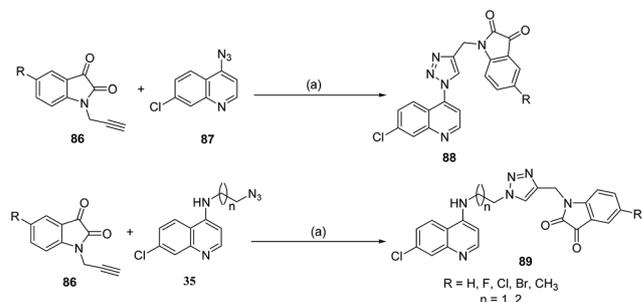
Fig. 10 General structures of synthesized 3-methylene-substituted indolinone analogues 80–85.

values of 11.7 and 13.5 nM, against the W2 resistant strain of *P. falciparum*.⁸⁸

2.9 Organometallic based 4-aminoquinoline conjugates

Over the past few years, bio-organometallic chemistry has emerged as a quickly growing and maturing area that associates organometallic chemistry to medicine, biology and molecular biotechnology.⁸⁹ The replacement of organic ligands with metal complexes has the ability to enhance the activity of biological compounds along with potential advantages, such as the synthesis of stable metal complexes with predictable structures, the capability to tune ligand affinities in accordance with their electron transfer properties, substitution rates and reduction potentials along with effective biological targeting.⁹⁰ Among the implausible number and variety of roles that metals play in current medicine, cancer⁹¹ and malaria⁹² treatments are appreciated as possibly the most important application of metal-based drugs.

In malaria chemotherapy, resistance to established drugs may be overcome by bio-organometallics through new metal specific modes of action. Ferroquine (99, FQ, SSR97193, Fig. 14), the ferrocene analogue of CQ, is the most pertinent example of a bio-organometallic compound that showed higher antimalarial



Scheme 15 Reagents and conditions: (a) CuSO_4 , sodium ascorbate, $\text{EtOH} : \text{H}_2\text{O}$, rt 7 h.

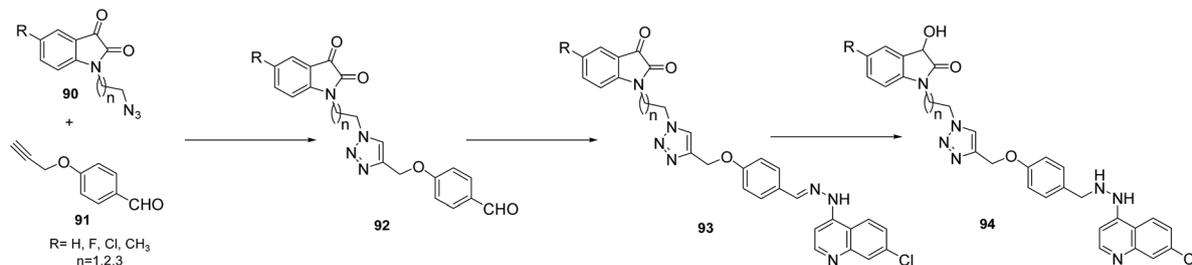


Fig. 11 General structures of target hybrid compounds.

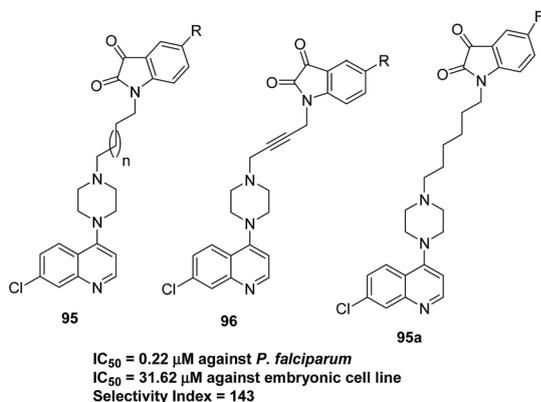


Fig. 12 General structures of piperazine tethered 7-chloroquinoline-isatin conjugates 95 and 96 along with most potent conjugate 95a.

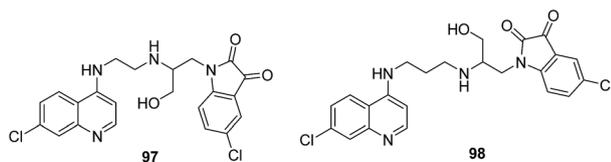


Fig. 13 The two most potent β-amino alcohol tethered isatin-4-aminoquinoline conjugates 97 and 98.

potency *in vivo* on mice infected with *P. yoelii* NS and *P. berghei* N. It is twenty two times more active against schizontocides than CQ against the CQ-resistant strain of *P. falciparum*. The effectiveness of FQ against the CQ-resistant strain could possibly be due to its capability to penetrate in the infected cells.⁹³

Biot *et al.* further utilized the ferroquine pharmacophore as a template for the design of two novel series of ferrocenic anti-malarial conjugates comprising FQ derivatives hybridized with

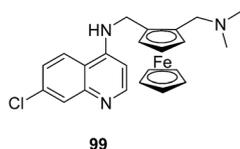
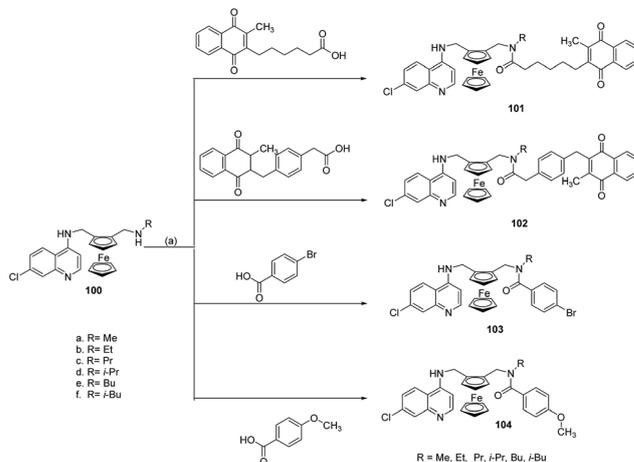


Fig. 14 Structure of ferroquine (FQ), 99.

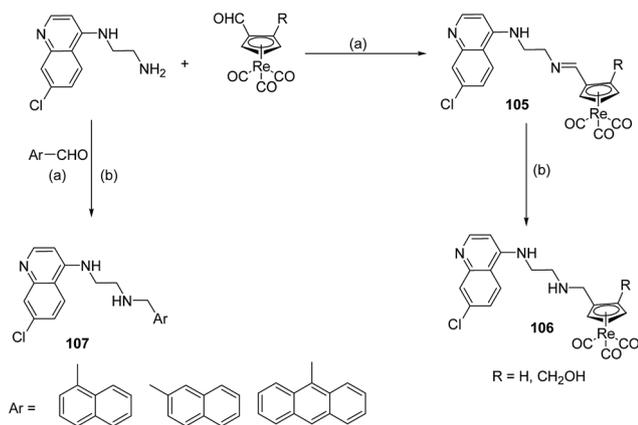
a glutathione reductase inhibitor (or glutathione depletor) *via* an appropriate cleavable amide bond (Scheme 16). *In vitro* antimalarial studies on CQ-susceptible (NF54) and CQ-resistant (K1) strains were conducted, with emphasis on the study of their mechanism of action.⁹⁴ Among the series of synthesized conjugates, the compounds 101a–101f were found to exhibit the most potent *in vitro* activity with IC₅₀ values in the nanomolar range. However, their antiplasmodial activity was slightly less than the precursor FQs 100a–100f possibly due to the cleavage of the amide bond and the oxidative metabolism of the side chain of the FQ in the digestive vacuole of the parasite. The observed antiplasmodial activity of FQ was mainly attributed to the high potential of the ferrocene core exhibiting a redox mechanism different from the mechanism of action of chloroquine.

The discovery of ferroquine (99, FQ, SSR97193) as a promising antimalarial prompted the synthesis of corresponding rhenium organometallics to measure the significance of the ferrocenyl group (Scheme 17).⁹⁵ All the synthesized compounds (organic along with their organometallic counterparts) were assessed for their *in vitro* antiplasmodial potency against the CQ-resistant (W2) and the CQ-susceptible (3D7) strains of *P. falciparum*. The cyrhetrene conjugates (105 and 106), however, were found to be less active than the corresponding ferrocene and organic analogues.

Orvig *et al.* explored the synthesis of CQ-bridged ferrocenophane analogues of ferroquine in which the terminal nitrogen



Scheme 16 Reagents and conditions: (a) EDCI, DCM, rt.



Scheme 17 Reagents and conditions: (a) MeOH, rt; (b) NaBH₄, MeOH, rt.

of the CQ derivative bridged the two cyclopentadienyl rings of ferrocene (**110a–110e**) alongside their mono-substituted ferrocene analogues (**109a–109e**) and organic fragments (**108a–108e**) (Fig. 15).⁹⁶ All the synthesized compounds were screened for their antiplasmodial potency and were found to be active over both CQ-sensitive and -resistant strains. Mono-substituted ferrocenyl analogues were found to be more active than the CQ-bridged ferrocenyl derivatives while the later compounds, *viz.* **110a–110e**, retained their activity against the CQ-resistant strains. SAR studies revealed the loss of activity with increased branching while the length of alkyl spacer did not seem to influence their antiplasmodial profiles. The physical properties, *viz.* the presence or absence of hydrogen bonding interactions, the degree of rigidity and the lipophilicity of these conjugates, were correlated to their biological activity. A balance between lipophilicity and hydrophilicity seemed to influence the biological profile of the conjugates while the structural characteristics, such as conformation and size, were found to be essential to overcoming the drug resistance.

The methodology was further extended with the synthesis of a new class of organometallic antimalarials comprising a ferrocene nucleus tethered to a CQ analogue and a 1,2,3,5-(diisopropylidene)- α -D-glucofuranose moiety in a 1,1'-heteroannular substitution pattern. Synthesis of these compounds was facilitated by orthogonal functionalization of ferrocene resulting in 1-acetoxy-1'-(1,3-dioxan-2-yl)ferrocene as a key intermediate for the modular introduction of the carbohydrate. This was followed by reductive amination with CQ derivatives, yielding 1-[3-(7-chloroquinolin-4-ylamino)alkylamino]-1'-[6-(1,2,3,5-diisopropylidene)- α -D-glucofuranosidyl]ferrocenes **117–119** (Fig. 16). These trifunctional hybrids were assessed for their antiplasmodial activity against CQ-susceptible (D10) and CQ-resistant (Dd2 and K1) strains of *P. falciparum*.⁹⁷ No correlation could be established between either the strength of intramolecular H-bonding or the polar surface area and anti-malarial potential of these compounds. Antiplasmodial studies indicated that the activity of the synthesized compounds (**117–119**) showed improvement in comparison to their either 1,2-disubstituted regio-isomers (**114–116**) or to

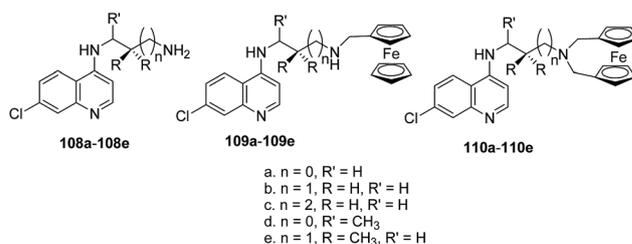


Fig. 15 CQ derivatives **108a–108e**, mono-substituted CQ–ferrocenyl compounds **109a–109e**, and 1,1'-disubstituted bridged CQ–ferrocenyl hybrids **110a–110e**.

monosubstituted ferrocenyls (**111–113**). The side chain length linking CQ with the ferrocene molecule did not have any effect on the observed activity.

After the discovery of FQ as a potential antimalarial drug, an extensive study was carried out to explore its derivatives with better activity, enhanced ADME properties and structure activity relationship. In a recent study, 4-aminoquinoline, trioxane and ferrocene were covalently linked to form a hybrid (**120**, Fig. 17) with the aim of studying its antimalarial efficacy. The scaffold revealed potent activity against CQ-resistant *P. falciparum* and was further evaluated *in vivo* against *P. vinckei petteri* infected mice. *In vivo* studies showed a marked decrease in the parasitemia to a lower than detectable level.⁹⁸ Two new “half sandwich” (η^6 -arenequinoline) Cr(CO)₃ complexes, *viz.* [η^6 -N-(7-chloroquinolin-4-yl)-N'-(2-dimethylaminomethylbenzyl)ethane-1,2-diamine]tricarbonylchromium and [η^6 -N-(7-chloroquinolin-4-yl)-N'-(2-dimethylaminobenzyl)ethane-1,2-diamine]tricarbonylchromium compounds (**121** and **122**, Fig. 17) were prepared and tested *in vitro* for their antiplasmodial efficacy against CQ-susceptible and CQ-resistant strains. The efficacy of complex **122** was twofold higher compared to the organic ligand.⁹⁹ In search of new efficient antimalarial complexes, Smith *et al.* synthesized a series of poly(propylene-imine) dendrimers functionalized with ferrocenyl thiosemicarbazones hybridized to the periphery of the molecule (**123**, Fig. 17). These metallo-dendrimers exhibited antiplasmodial activity in the low μ M range against the W2 strain of *P. falciparum*. Their antiplasmodial potencies were better than non-conjugated ferrocenyl thioesters as well as the free dendritic ligand.¹⁰⁰ Further, organometallic complexes, *viz.* cymantrene (CpMn(CO)₃) and cyrhetrene

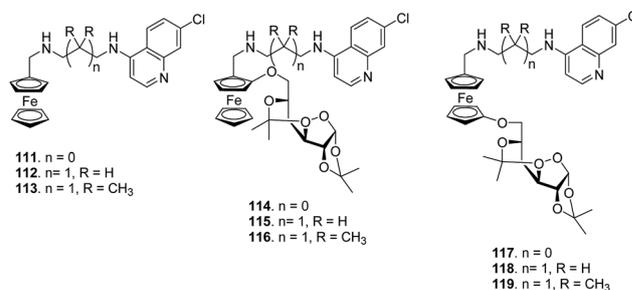


Fig. 16 Mono-, 1,2-disubstituted, 1,1'-disubstituted ferrocenyl chloquinone glucofuranose conjugates **111–113**, **114–116** and **117–119**.

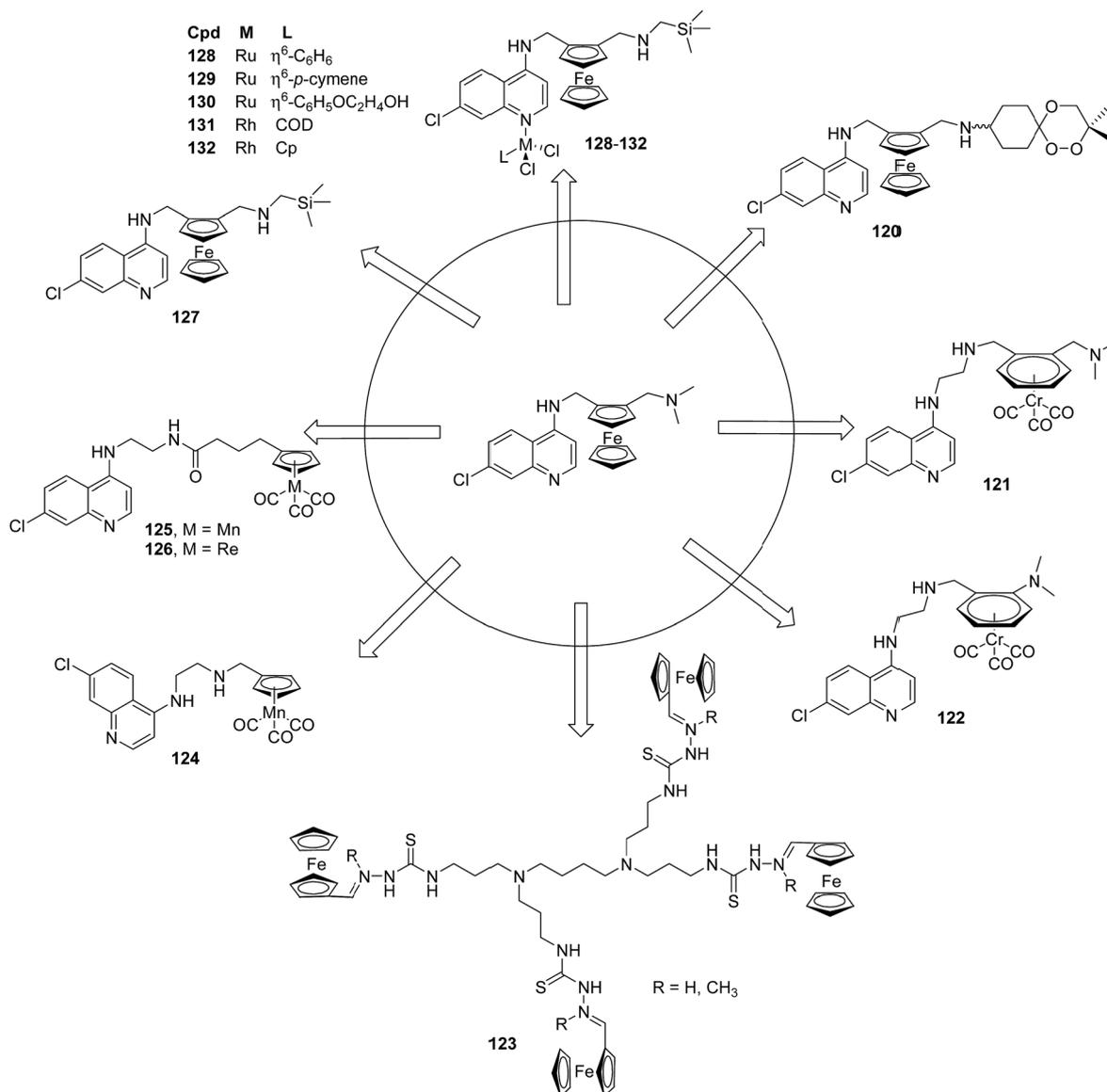


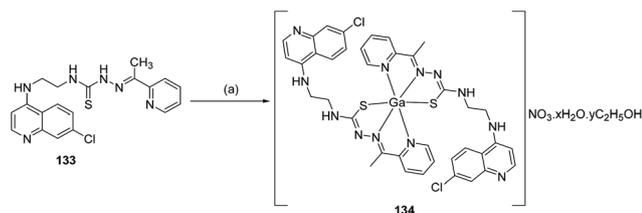
Fig. 17 Some conjugates of ferriquinoline.

(CpRe(CO)₃) tethered with 4-aminoquinolines, were reported (124–126, Fig. 17) and bio-evaluated for their malarial potency against D10 and Dd2 strains of *P. falciparum*. The low resistance indices (RI) of these complexes indicated that these complexes of manganese and ruthenium may act as a lead for further development of new antimalarial compounds.¹⁰¹

Smith *et al.* in a recent communication synthesized a new carbosilane congener of ferriquinoline (127, Fig. 17) by including an organosilicon moiety in the side chain of ferriquinoline. The carbosilane congener was further utilized for the synthesis of a corresponding series of metal complexes of neutral heterometallic ruthenium and rhodium (128–132), as shown in Fig. 17. All the newly synthesized compounds were evaluated for antiplasmodial activity against the NF54 and Dd2 strains of *P. falciparum*. The presence of the trimethylsilyl group in compound 127 exhibited superior activity (IC₅₀ = 7.32 nM) as

compared to ferriquinoline (IC₅₀ = 42.65 nM) against the NF54 strain. Among synthesized complexes, 130 was the most active with IC₅₀ values of 4.86 and 35.91 nM against NF54 and Dd2 strains, respectively.¹⁰²

Kumar and co-workers reported the synthesis of a gallium(III) complex with 7-chloroquinoline thiosemicarbazone as a ligand with antimalarial efficacy against 3D7 strain of *P. falciparum* (Scheme 18). The synthesized complex 134 exhibited improved antimalarial activity than lumefantrine on 3D7 isolate of *P. falciparum*. Further, the synthesized complex 134 was evaluated for anti-proliferative activity against different cancer cell lines and complex proved to be 31 times more potent on colon cancer cell line HCT-116 compared with the standard drug etoposide with considerably less cytotoxicity on non-cancerous colon fibroblast CCD-18Co.¹⁰³



Scheme 18 Reagents and conditions: (a) $\text{Ga}(\text{NO}_3)_3 \cdot \text{H}_2\text{O}$, ethanol, reflux, 2 h.

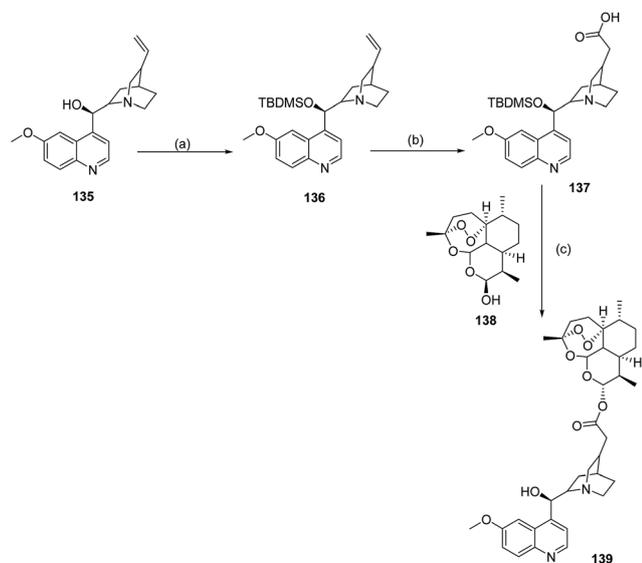
2.10 Artemisinin and trioxaquinone based 4-aminoquinoline conjugates

The benefits of molecular hybridization over combination therapy can be well rationalized by the report of Walsh and co-workers involving the synthesis of dihydroartemisinin–quinine hybrids (**139**). The compound was prepared by following a sequence of steps as shown in Scheme 19 involving an initial protection of quinine with TBDMSiCl with subsequent oxidation to form carboxylic acid and coupling with dihydroartemisinin.⁴⁵ The antimalarial efficacy of the synthesized hybrids was determined against CQ-sensitive 3D7 and CQ-resistant FcB1 strains of *P. falciparum*. The conjugate proved to be more potent than either quinine or dihydroartemisinin while a three-fold improvement in efficacy was observed over a 1 : 1 mixture of quinine and dihydroartemisinin.

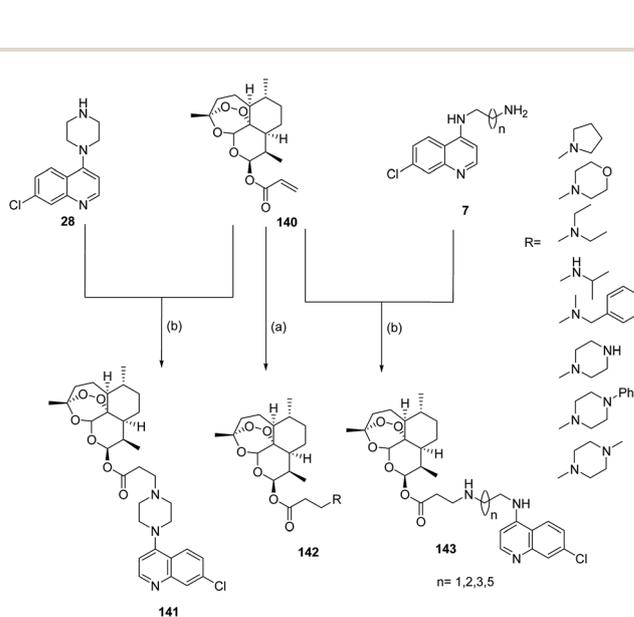
Rationalization of literature has revealed that the variation in the length of alkyl chain between two nitrogen atoms of the 4-diaminoalkyl side chain has a marked influence on the antimalarial efficacy. The compounds with either smaller alkyl chain length ($n = 2-4$) or with longer alkyl chain length ($n = 10-12$) retained their activities against CQ-resistant strains,

whereas the compounds with chain length $n = 5-8$ showed a relative decrease in the activity.¹⁰⁴ Chibale and co-workers exploited a molecular hybridization protocol for the preparation of dihydroartemisinin–7-chloroquinoline conjugates (**141** and **143**) prepared *via* aza-Michael addition reaction along with the analogues of dihydroartemisinin (DHA) derivatives **142** (Scheme 20). These were tested for their antimalarial profiles against CQ-sensitive (D10) and CQ-resistant (Dd2) strains of *P. falciparum*. The length of alkyl chain introduced as the spacer was chosen to be between 2 and 6 carbon atoms for optimal biological activity.¹⁰⁵ The evaluation data revealed a substantial increase in antimalarial efficacy of the hybrids over the precursors **7** or **28** while a decrease in antimalarial efficacy was observed with the increase in chain length. DHA derivatives prepared *via* aza-Michael addition with available amines were found to be more effective in the growth inhibition of *P. falciparum* suggestive of the fact that the DHA nucleus is mainly responsible for the observed antimalarial activity of the hybrids. The target compound **141** and three dihydroartemisinin derivatives of **125** showed excellent antiplasmodial activity with IC_{50} values of ≤ 10 nM against both D10 and Dd2 strains of *P. falciparum* and low cytotoxicity against human cell lines (HeLa cells).

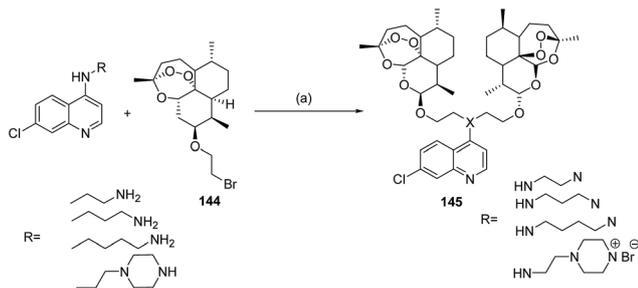
N'Da further extended his work on the synthesis of artemisinin–quinolines and their hybrid-dimers from dihydroartemisinin and aminoquinolines (Scheme 21).¹⁰⁶ The synthesized hybrids (**145**) obtained as β -isomers were assessed for their antiplasmodial activity against CQ-sensitive and -resistant strains of *P. falciparum* along with cytotoxicity profiles against a mammalian cell-line. Although none of the hybrids was found to be more potent than DHA, two were more active than CQ. A hybrid-dimer with an optimum chain length of three carbon atoms displayed excellent antimalarial potency, with IC_{50} values of 5.31 and 28.43 nM against D10 and Dd2 strains of *P. falciparum*.



Scheme 19 Reagents and conditions: (a) TBDMSiCl, Et_3N , DMAP, DMF, rt; (b) (i) BH_3 -THF, diglyme, 0°C ; (ii) $\text{Me}_3\text{NO} \cdot 2\text{H}_2\text{O}$, 100°C ; (iii) Jones reagent, acetone, rt; (c) (i) 2,6-dichlorobenzoyl chloride, Et_3N , DMAP, DCM, rt; (ii) TBAF, THF, rt.



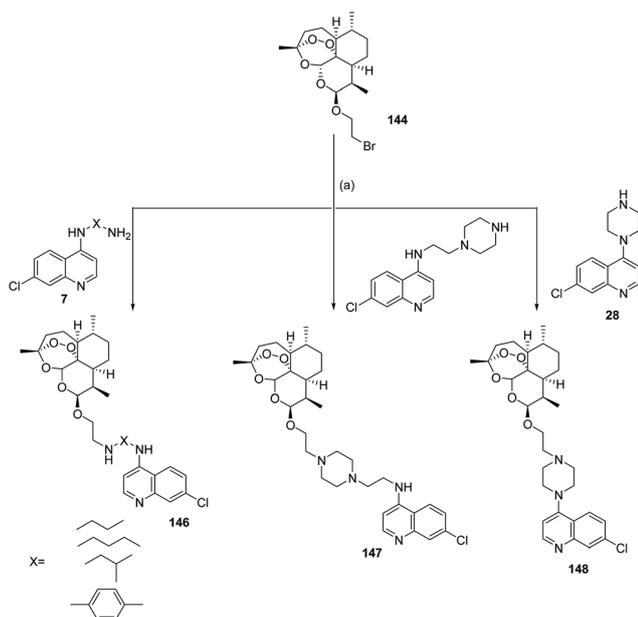
Scheme 20 Reagents and conditions: (a) appropriate amine, DBU, CH_3CN , rt, 12 h; (b) DBU, DMF, N_2 , rt, 12 h.



Scheme 21 Reagents and conditions: (a) DMF, 90–110 °C, 6–8 h.

N'Da with his co-workers explored the synthesis of dihydroartemisinin-4-aminoquinoline conjugates (**146**, **147** and **148**) with their subsequent conversion to oxalate salts (Scheme 22). *In vitro* antimalarial activity was tested against CQ-sensitive D10 and -resistant Dd2 strains of *P. falciparum* while cytotoxicity was determined against mammalian Chinese Hamster Ovarian (CHO) *via* 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT)-assay.¹⁰⁷ Most of the synthesized conjugates showed comparable to higher activity to that of the standard drug CQ with IC_{50s} ranging from 17.2 to 38.9 nM. The oxalate salts proved to be seven times more potent than the standard drug CQ against the CQ-resistant strain of *P. falciparum*.

In 2011, Chibale *et al.* extended the synthesis of artemisinin-4-aminoquinoline conjugates (**151**, **152**) *via* Ugi four-component condensation (Ugi-4CR) reaction (Scheme 23)¹⁰⁸ using paraformaldehyde as aldehyde component, 7-chloro-4-diaminoalkylquinoline as the amine input, artelinic acid/1,4-naphthoquinone acid as acid input, and cyclohexyl- or *tert*-butyl isocyanide as the isocyanide input. Antiplasmodial studies of the synthesized conjugates showed comparable potency against CQ-resistant K1 and CQ-sensitive D10 strains of *P. falciparum*.



Scheme 22 Reagents and conditions: (a) DMF, 70–80 °C, 4–6 h.

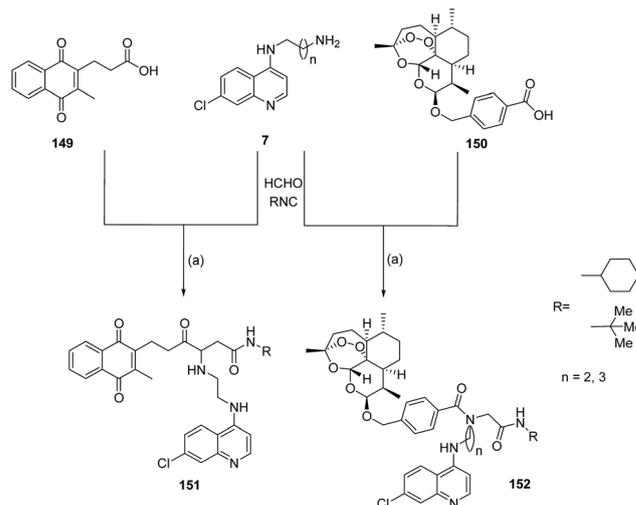
Mechanistically, the conjugates exhibited potent inhibitory activities of β -hematin formation and appeared to inhibit endocytosis as evident by the decrease in the number of transport vesicles in the parasite.

Meunier *et al.* reported the synthesis of trioxaquinones **154** as potential antimalarials by following the synthetic protocol as depicted in Scheme 24.¹⁰⁹ Compound trioxaquinone (PA1103/SAR116242) **154**, obtained from this study, proved to be highly active against several sensitive and resistant strains of *P. falciparum* at nM concentrations. It has also shown activity on multidrug-resistant strains attained from fresh patient isolates in Gabon along with good drug profiles *viz.* absorption, metabolism and safety parameters. Efficacy of this conjugate *via* oral route is very high with total curing of mice infected with CQ-sensitive and -resistant strains of *Plasmodium* at 26–32 mg kg⁻¹.

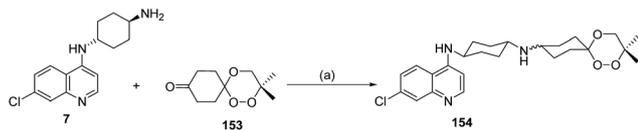
The selection of trioxaquinone (PA1103/SAR116242) for anti-malarial drug development strongly confirms the role of molecular hybridization in furnishing molecules with satisfactory pharmacological and safety profiles to allow regulatory drug development. Guided by similar rationale, O'Neill and co-workers reported the synthesis of semi-synthetic trioxaquinones and trioxolaquinones in excellent yields and assessed their anti-malarial activities against CQ-sensitive (3D7) and -resistant (K1) strain of *P. falciparum*. Compounds of both the series showed activity in low nM range with some of the analogues being more active than artemisinin. The structure of two of the most active compounds **155** and **156** of the series is as shown in Fig. 18.¹¹⁰

2.11 Miscellaneous antimalarial 4-aminoquinoline conjugates

A library of tetrahydro-1*H*- β -carboline-4-aminoquinoline hybrids **159** were identified as a novel class of highly potent antimalarial agents by Chauhan *et al.* using molecular hybridization approach with MIC values ranging from 0.05 to 19.08 μ M against CQ-sensitive strain NF-54 of *P. falciparum* (Scheme 25).¹¹¹ Most potent of the synthesized conjugate was



Scheme 23 Reagents and conditions: (a) succinic acid, silver nitrate, ammonium persulfate, 30% aq. CH₃NH₂, 65–70 °C, 3 h.



Scheme 24 Reagents and conditions: (a) NaBH_3CN , MeOH, HCl/i-PrOH.

found to have seven folds more activity than the standard drug CQ and therefore warrants the development of these hybrids in antimalarial chemotherapy.

In 2011, Egan and collaborators proposed the synthesis of dibemethin-coupled 4-aminoquinolines **162** as potential antimalarial agents with a view to obtain novel hybrids capable of inhibition of hemozoin formation in the digestive vacuole of parasite (Scheme 26).¹¹² These hybrids were equally potent against cultured CQ-sensitive (D10) and CQ-resistant (K1) *P. falciparum*. Several compounds exhibited *in vitro* antimalarial potency below 100 nM against both strains of the parasite while none of compound exhibited cross-resistance with the standard drug CQ. The compounds with promising *in vitro* activity and non-cytotoxicity against mammalian cell line were screened for their *in vivo* activity against *P. berghei*. The active compounds exhibited significant *in vivo* activity and reduces parasitemia by over 99%. SAR studies showed marked dependence of activity on the site of attachment of 4-aminoquinoline with side chain of dibemethin as well as the substituent on the terminal phenyl ring of dibemethin. Researchers further investigated the interaction of this series of compounds with CQ transport by making use of the *Xenopus* oocyte expression system, which directly measures CQ transport by *P. falciparum* CQ-resistance transporter (PfCRT) and its inhibition with potential resistance-reversers or reversed-CQ compounds. The results of this study confirmed 4-aminoquinoline-dibemethin hybrids as the first example of reversed-CQ antimalarials with capability to inhibit both PfCRT and hemozoin formation.

Recently, Chibale *et al.* reported the design and synthesis of novel β -amino alcohol derivatives of 4-aminoquinoline (**164**) with an extension to 4-aminoquinoline-oxazolidinone conjugates (**165**, Scheme 27).¹¹³ *In vitro* antimalarial activity was evaluated against CQ-sensitive (3D7) and CQ-resistant (K1) strains of *P. falciparum*. Few of the β -amino alcohol analogues

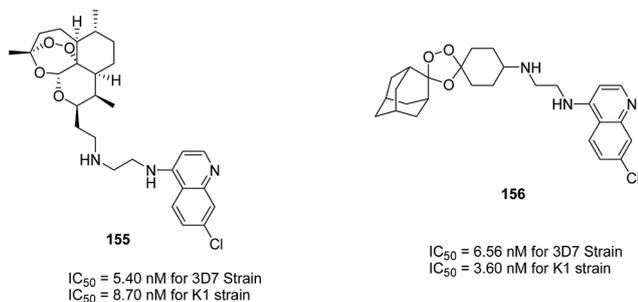
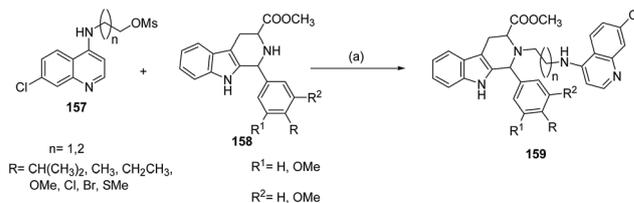


Fig. 18 Most potent 1,2,4-trioxaquines and 1,2,4-trioxolaquines conjugates.

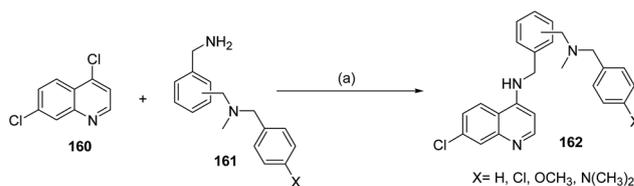


Scheme 25 Reagents and conditions: (a) DMF, 120 °C, high pressure.

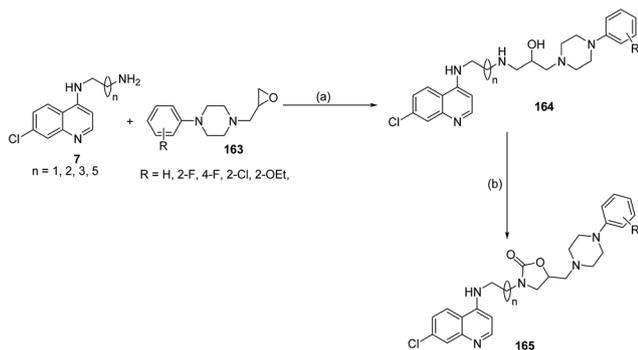
were more active compared to CQ against the CQ-sensitive strain while the potency decreased considerably against the CQ-resistant strain, revealing cross resistance of these conjugates with CQ. The conversion of β -amino alcohol to oxazolidinones by reacting with tris-phosgene resulted in a considerable decrease in the activities of the synthesized conjugates against both CQ-sensitive and CQ-resistant strains.

Chauhan *et al.* synthesized sixteen new 4-aminoquinoline analogues by modifying its side chain with an extension towards the synthesis of quinoline-acridine conjugates and tested their *in vitro* antiplasmodial activity against the NF 54 strain of *P. falciparum*.¹¹⁴ Among the tested compounds, the compound **166** ($\text{MIC} = 0.125 \mu\text{g ml}^{-1}$) was nearly as active as CQ ($\text{MIC} = 0.125 \mu\text{g ml}^{-1}$) while another (**167**; $\text{MIC} = 0.031 \mu\text{g ml}^{-1}$) exhibited four-fold more potency than CQ. Three of the selected test compounds (**166**, **167** and **168**; Fig. 19) with good *in vitro* antiplasmodial potency were chosen for *in vivo* screening in Swiss mice infected with the CQ-resistant N-67 strain of *P. yoelii*. Compound **167** showed a curative response to all treated Swiss mice at doses of 50 mg kg^{-1} and 25 mg kg^{-1} for 4 days given *via* intra-peritoneal route. The compound was orally active at the dose of 100 mg kg^{-1} and has proved the importance of exploring the 4-aminoquinoline class for the discovery of new antimalarial agents.

The hybrid design strategy was further exemplified by Whitlock and co-workers in the synthesis of CQ-astemizole (non-sedating H1 antagonist) conjugates **170** along with their antiplasmodial evaluation against K1 strain of *P. falciparum* (Fig. 20).¹¹⁵ Most of the conjugates were three to four folds more potent than CQ with the most potent of the tested compound exhibiting an IC_{50} of 23 nM. Importantly, the compounds displayed more than 100 times selectivity for antiparasitic activity over cell-based cytotoxicity. Based on the preliminary *in vitro* *P. falciparum* activity, two potent compounds were further screened for *in vivo* studies on a *P. berghei* mouse malaria model.



Scheme 26 Reagents and conditions: (a) anhyd. *N*-methyl-2-pyrrolidine, K_2CO_3 , Et_3N .



Scheme 27 Reagents and conditions: (a) MeOH, 55 °C; (b) tris-phosgene, DCM, 0 °C-rt.

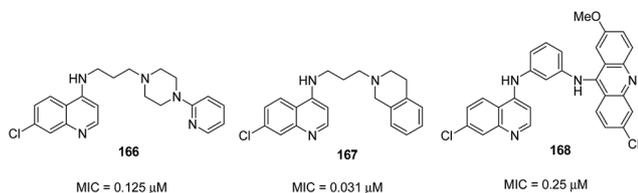


Fig. 19 Most potent compounds among the series.

Santos and colleagues synthesized molecular hybrids of commercially available squarates with the antimalarial drugs, chloroquine (CQ) and primaquine (PM) with improved antiplasmodial activity compared with the squarates.¹¹⁶ Three of the synthesized conjugates containing two 7-chloroquinoline moieties were two fold more active than the standard drug, CQ with IC_{50} values ranging from 0.095–0.20 μ M against the CQ-resistant W2 strain of *P. falciparum* (Fig. 21). The length of the carbon chain introduced as the linker between the squarates and quinoline scaffolds has been shown to have a major impact on the observed activity profiles with the activity generally increasing with the increase in chain length.

Vacuolar plasmepsins, pepsin-like aspartic proteases present in different species of the parasite *Plasmodium*, are recognized as promising targets for the enlargement of new antimalarial agents.¹¹⁷ They comprise plasmepsins 1 (PLM 1), plasmepsins 2 (PLM 2), plasmepsins 4 (PLM 4) and a histo-aspartic protease

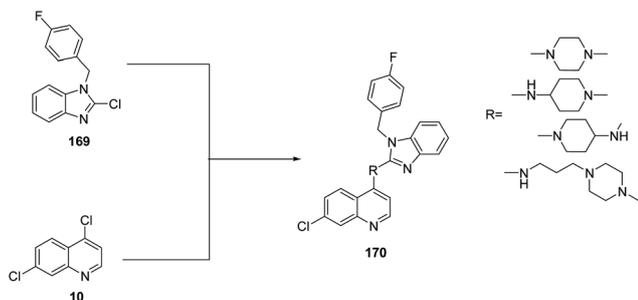


Fig. 20 General strategy of 7-chloroquinoline–astemizole based hybrids.

(HAP) and play a vital role in the survival of the parasite. These enzymes are contained in the food vacuole of the parasite and are directly involved in the degradation of human haemoglobin during the erythrocytic stage. Statin-based molecules are shown to be potent inhibitors of PLMs, however with restricted effectiveness in killing the parasite (IC_{50} range 2–20 μ M).¹¹⁷ In 2012, Romeo and co-workers, in an attempt to circumvent the limitations associated with statin-based molecules, synthesized its hybrids with 4-aminoquinoline 172.¹¹⁸ It was considered that since both of these molecules interfere with the haemoglobin catabolism process, the resulting hybrid would have an improvement in overall antimalarial potency.¹¹⁹ Two of the 4-aminoquinoline–statin hybrids (172a and 172b) were more potent than CQ against the CQ-resistant strain (Fig. 22) with good selectivity against cathepsin D. It was concluded from β -hematin inhibitory assay that the 7-chloro group in the quinoline ring is obligatory for the antiplasmodial efficacy of the synthesized conjugates.

Chibale and co-workers reported the preparation of 4-aminoquinoline–3-hydroxypyridin-4-one conjugates on the basis of additive *in vitro* combination of *N*-alkyl-3-hydroxypyridin-4-one with chloroquine (CQ) (Scheme 28). All the synthesized conjugates were assessed *in vitro* against CQ-resistant (K1 and W2) and -sensitive (3D7) strains of *P. falciparum* along with β -hematin formation studies.¹²⁰ Most of the conjugates were shown to have superior β -hematin inhibition activity to that of CQ and a strong correlation was observed between antimalarial profiles and inhibition of β -hematin formation. Among the synthesized conjugates, three potent compounds with longer alkyl chains ($n = 3, 5$) against K1, 3D7, and W2, respectively, having IC_{50} s of 175c (0.13, 0.004, and 0.1 μ M); 175d (0.08, 0.01, and 0.02 μ M); and 174g (0.07, 0.03, and 0.08 μ M). A cytotoxicity study on a mammalian cell line showed that most of the conjugates screened were non-cytotoxic and far less toxic than podophyllotoxin (POD).

Another attempt to explore the antimalarial potential of bis-quinolines was made by N'Da and co-workers by exploring the synthesis of a series of bis-quinolines and bis-pyrrolo[1,2a]quinoxalines containing various polyamine linkers along with the determination of their aqueous solubility and distribution coefficients (Scheme 29).¹²¹ The antimalarial potency of these compounds was assessed against CQ-sensitive (D10) and CQ-resistant (Dd2) strains of *P. falciparum* besides the growth inhibitory studies against a panel of cancer cell lines. Bis-quinolines showed a general increase in antimalarial activity

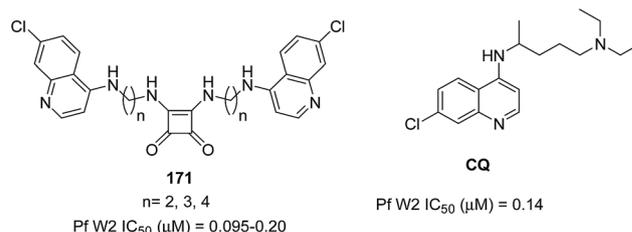


Fig. 21 Most potent 4-aminoquinoline–squarate hybrids and CQ.

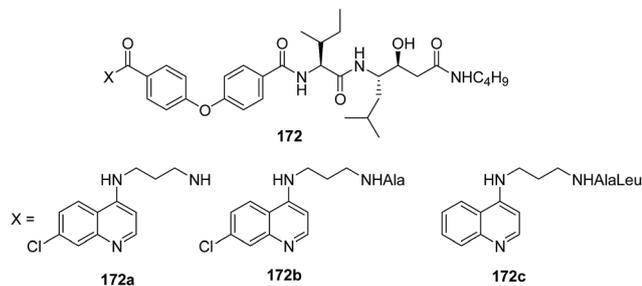
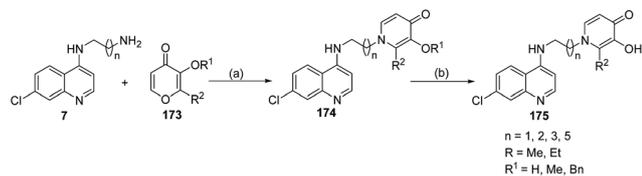


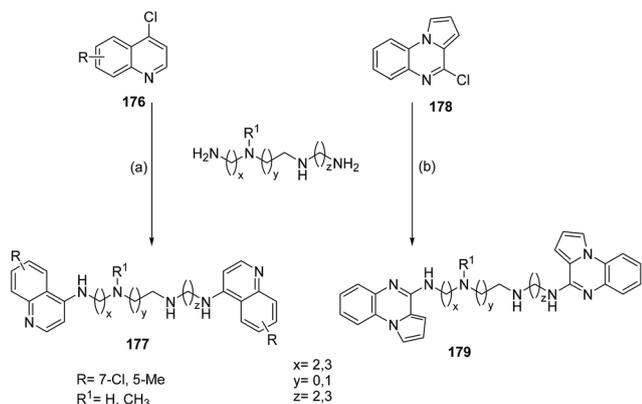
Fig. 22 Most potent 4-aminoquinoline–statin hybrids **171a**, **171b**, **171c**.



Scheme 28 Reagents and conditions: (a) 50% aq. EtOH, pH 13 (2 M NaOH), 90–120 °C, 18–24 h; (b) H₂, Pd/C, 2 M ethanolic HCl, 4 atm or 1 : 2 : 3 H₂O/EtOH/HCl, 74 °C, 24 h.

with the increase in protonation sites given by polyamine linkers that could help to overcome CQ-resistance. The 7-chloro-substituted hybrids have been shown to possess more potency compared to either 5-methyl quinoline or pyrrolo[1,2*a*] quinoxaline conjugates. Among the bis-quinoline, triethylene-tetramine and *N,N*-bis(3-aminopropyl)ethylene-diamine linkers were found to be the most potent of all the synthesized conjugates.

Chibale *et al.* explored the molecular hybridization paradigm for the synthesis of three groups of hybrid molecules by covalent fusion of azidothymidine (AZT) with dihydroartemisinin (DHA), a tetraoxane or a 4-aminoquinoline derivative that target *P. falciparum* and HIV simultaneously. All synthesized compounds were tested for their inhibitory activity against pseudotyped HIV-1, CQ-sensitive (3D7) and CQ-resistant (Dd2)



Scheme 29 Reagents and conditions: (a) 135–145 °C, 16–20 h; (b) K₂CO₃, DMF, 135–145 °C, 16 h.

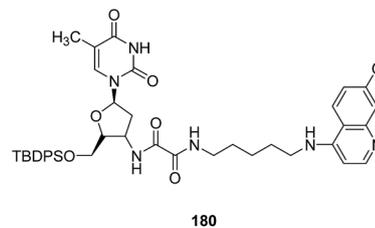


Fig. 23 Most potent CQ–AZT compound **180**.

strains of *P. falciparum* at three different concentrations (50, 5 and 1 μM) along with cytotoxicity assessment against HeLa cells. Among the tested conjugates, a potent CQ–AZT conjugate (**180**, Fig. 23) exhibited IC₅₀ values of 0.38 and 0.08 μM (more potent than CQ and tetraoxane) against 3D7 and Dd2 strains of *P. falciparum*, respectively. This compound also exhibited enhanced inhibitory activity (IC₅₀ value of 0.9 μM) against pseudotyped HIV-1 with a selective index of 31.8. However, higher cytotoxicity in HeLa cells discontinued its further testing.¹²²

In a recent communication, Rao *et al.* reported the synthesis of a series of 4-aminoquinoline–4*H*-chromene conjugates along with their antimalarial evaluation against two *P. falciparum* strains, namely 3D7 (CQ-sensitive) and K1 (CQ-resistant).¹²³ All synthesized conjugates possessed activity in the μM range with compounds **181** having piperazine linkage and nitro substituent and **182** with azapane linkage and chloro substituent at the C-6 position of 4*H*-chromene showing comparable activity to that of CQ (Fig. 24). Further, molecular docking suggested that these conjugates exhibited strong binding affinity with *P. falciparum* lactate dehydrogenase (PfLDH) and act as potent inhibitors of the parasite specific glycolytic pathway.

Chibale *et al.* recently disclosed the synthesis of 4-aminoquinoline–benzoxazole conjugates and screened for their anti-malarial efficacy against K1 (multidrug resistant) and NF54 (sensitive) strains of the parasite *P. falciparum*.¹²⁴ The anti-plasmodial activities of the synthesized compounds exhibited a good structure–activity relationship, which led to the recognition of highly promising conjugates having IC₅₀s in the nM range against both K1 and NF54 strains of *P. falciparum*. Further, the synthesized conjugates showed good *in vitro* microsomal metabolic stability along with desirable *in vivo*

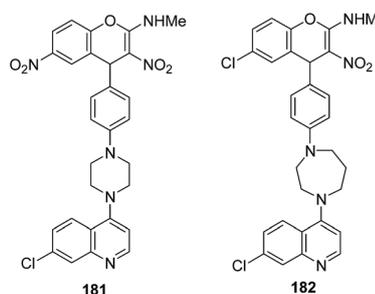


Fig. 24 Most potent 7-chloroquinoline–4*H*-chromene conjugates **181** and **182**.

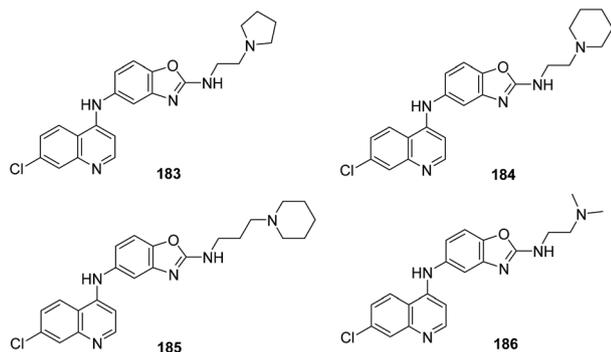
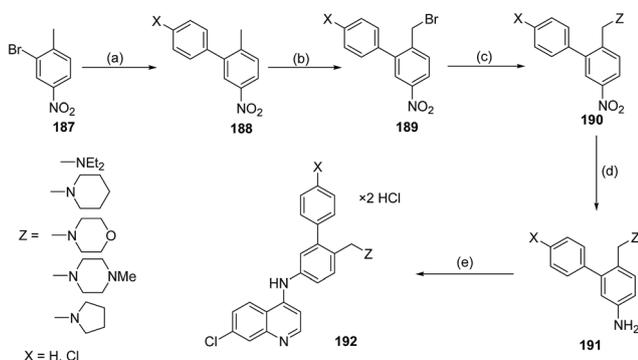


Fig. 25 Most potent 7-chloroquinoline–benzoxazole conjugates **183**, **184**, **185** and **186**.

pharmacokinetic results. The four most promising conjugates (**183**, **184**, **185** and **186**, Fig. 25) were subjected to *in vivo* antimalarial studies against *P. berghei* infected mice. Compound **183** showed good *in vivo* oral efficacy and completely cured the treated mice at a low multiple dose of 4–10 mg kg⁻¹. Mechanistic studies revealed the inhibition of hemozoin formation as one of the probable mechanisms of action for these compounds. Further, β -haematin inhibition studies confirmed the importance of the quinoline nucleus for antiplasmodial activity as non-benzoxazole intermediates did not show any inhibitory activity even at highest tested concentration.

Lopez and co-workers in a recent communication showed the synthesis of 3'-dehydroxy-isotebuquine analogs of 4-aminoquinoline **192** via a five step synthetic sequence in good to excellent yields as shown in Scheme 30. The synthesized conjugates were assessed for their β -hematin inhibitory activity. Among eight of the synthesized conjugates, six compounds were shown to have greater than 97% IHF value (% inhibition of hemozoin formation). These potent compounds were further tested for their *in vivo* antimalarials activity in mice infected with *P. berghei* ANKA CQ-susceptible strain with three of the conjugates displaying *in vivo* antimalarial efficacy comparable to that of CQ.¹²⁵



Scheme 30 Reagents and conditions: (a) phenylboronic acid, toluene, Pd(PPh₃)₄ (5%), K₂CO₃, EtOH : H₂O (1 : 1), 100 °C, 24 h; (b) NBS, CCl₄, (PhCO)₂O₂, light, reflux, 24 h; (c) dialkylamine, toluene, reflux, 6 h; (d) Sn/HCl, 70 °C, 2 h; (e) 4,7-dichloroquinoline, EtOH, HCl (cat.), reflux, 6 h.

3. Conclusions

The development of new drugs with improved physicochemical profiles, lack of toxicity, synthetic selectivity and economic accessibility represents a big challenge for the pharmaceutical sector and warrants continuous efforts. The present review establishes the attention of organic medicinal chemists in the field of 4-aminoquinoline-hybridization towards the synthesis of new molecular frameworks for averting and delaying the emergence of drug resistance along with the improvement of efficacy. The validation of molecular hybridization, however, is crucial and the benefit of the hybrid over the separate pharmacophores or their 1 : 1 combination should be confirmed.

Notes and references

- 1 T. Bishop and P. Sham, *Analysis of Multifactorial Diseases*, Academic Press, New York, 2000, pp. 1–320.
- 2 M. H. Cohen, G. A. Williams, R. Sridhara, G. Chen, W. D. J. McGuinn, D. Morse, S. Abraham, A. Rahman, C. Liang, R. Lostritto, A. Baird and R. Pazdur, *Clin. Cancer Res.*, 2004, **35**, 1212.
- 3 S. Frantz, *Nature*, 2005, **437**, 942.
- 4 H. Takano, K. Sawada, N. Sato, A. Natoya, T. Tarumi, S. Hirayama, K. Koizumi, T. A. Takahashi, S. Sekiguchi and T. Koike, *Leuk. Lymphoma*, 1996, **21**, 473.
- 5 B. A. Larder, S. D. Kemp and P. R. Harrigan, *Science*, 1995, **269**, 696.
- 6 S. A. Eisen, D. K. Miller, R. S. Woodward, E. Spitznagel and T. R. Przybeck, *Arch. Intern. Med.*, 1990, **150**, 1881.
- 7 N. S. Skolnik, J. D. Beck and M. Clark, *Am. Fam. Physician*, 2000, **61**, 3049.
- 8 G. Glass, *Nat. Rev. Drug Discovery*, 2004, **3**, 731.
- 9 W. H. Frishman and A. L. Zuckerman, *Expert Rev. Cardiovasc. Ther.*, 2004, **2**, 675.
- 10 N. A. Flores, *Curr. Opin. Invest. Drugs*, 2004, **5**, 984.
- 11 T. N. C. Wells, P. L. Alonso and W. E. Gutteridge, *Nat. Rev. Drug Discovery*, 2009, **8**, 879.
- 12 P. G. Clay, T. A. H. Taylor, A. G. Glaros, M. P. McRae, C. Williams, D. McCandless and M. Oelklaus, *Ther. Clin. Risk Manage.*, 2008, **4**, 291.
- 13 R. Morphy and Z. Rankovic, *J. Med. Chem.*, 2005, **48**, 6523.
- 14 B. Meunier, *Acc. Chem. Res.*, 2008, **4**, 69.
- 15 F. W. Muregi and A. Ishih, *Drug Dev. Res.*, 2010, **71**, 20.
- 16 C. Viegas-Junior, A. Danuello, V. D. S. Bolzani, E. J. Barreiro and C. A. M. Fraga, *Curr. Med. Chem.*, 2007, **14**, 1829.
- 17 C. A. M. Fraga, *Expert Opin. Drug Discovery*, 2009, **4**, 605.
- 18 R. Morphy and Z. Rankovic, *Curr. Pharm. Des.*, 2009, **15**, 587.
- 19 J. O. Gyapong, M. Gyapong, N. Yellu, K. Anakwah, G. Amofah, M. Bockarie and S. Adjiei, *Lancet*, 2010, **375**, 160.
- 20 World Health Organization, *Global report for research on infectious diseases of poverty*, 2012, http://whqlibdoc.who.int/publications/2012/9789241564489_eng.pdf.

- 21 WHO, *World Malaria Report*, 2013, http://www.who.int/malaria/publications/world_malaria_report_2013/en/index.html.
- 22 R. Banerjee, J. Liu, W. Beatty, L. Pelosof, M. Klemba and D. E. Goldberg, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 990.
- 23 N. D. Gamboa de Domínguez and P. J. Rosenthal, *Blood*, 1996, **87**, 4448.
- 24 P. Loria, S. Miller, M. Foley and L. Tilley, *Biochem. J.*, 1999, **339**, 363.
- 25 C. D. Fitch, R. Chevli, P. Kanjananggulpan, P. Dutta, K. Chevli and A. C. Chou, *Blood*, 1983, **62**, 1165.
- 26 T. Yoshida and T. C. Migita, *J. Inorg. Biochem.*, 2000, **82**, 33.
- 27 K. A. de Villiers and T. J. Egan, *Molecules*, 2009, **14**, 2868.
- 28 T. J. Egan and H. M. Marques, *Coord. Chem. Rev.*, 1999, **190**, 493.
- 29 T. J. Egan, R. Hunter, C. H. Kaschula, H. M. Marques, A. Misplon and J. Walden, *J. Med. Chem.*, 2000, **43**, 283.
- 30 M. Schlitzer, *Curr. Med. Chem.*, 2007, **2**, 944.
- 31 V. Narasimhan and A. Attaran, *Malar. J.*, 2003, **2**, 8.
- 32 T. E. Wellems, *Science*, 2002, **298**, 124.
- 33 A. B. S. Sidhu, D. Verdier-Pinard and D. A. Fidock, *Science*, 2002, **298**, 210.
- 34 G. Edwards and G. A. Biagini, *Br. J. Clin. Pharmacol.*, 2006, **61**, 690.
- 35 T. K. Mutabingwa, *Acta Trop.*, 2005, **5**, 305.
- 36 R. T. Eastman and D. A. Fidock, *Nat. Rev. Microbiol.*, 2009, **7**, 864.
- 37 A. M. Dondorp, F. Nosten, P. Yi, D. Das, A. P. Phyto, J. Tarning, K. M. Lwin, F. Ariey, W. Hanpithakpong, S. J. Lee, P. Ringwald, K. Silamut, M. Imwong, K. Chotivanich, P. Lim, T. Herdman, S. S. An, S. Yeung, P. Singhasivanon, N. P. Day, N. Lindegardh, D. Soheat and N. J. White, *N. Engl. J. Med.*, 2009, **361**, 455.
- 38 O. Dechy-Cabaret, F. Benoit-Vical, C. Loup, A. Robert, H. Gornitzka, A. Bonhoure, H. Vial, J. F. Magnaval, J. P. Seguela and B. Meunier, *Chem.-Eur. J.*, 2004, **10**, 1625.
- 39 F. Bellot, F. Cosledan, L. Vendier, J. Brocard, B. Meunier and A. Robert, *J. Med. Chem.*, 2010, **53**, 4103.
- 40 A. Kumar, K. Srivastava, S. R. Kumar, M. I. Siddiqi, S. K. Puri, J. K. Saxena and P. M. S. Chauhan, *Eur. J. Med. Chem.*, 2011, **46**, 676.
- 41 S. Manohar, M. Tripathi and D. S. Rawat, *Curr. Top. Med. Chem.*, 2014, **14**, 1706.
- 42 E. M. Guantai, K. Ncokazi, T. J. Egan, J. Gut, P. J. Rosenthal, R. Bhampidipati, A. Kopinathan, P. J. Smith and K. Chibale, *J. Med. Chem.*, 2011, **54**, 3637.
- 43 F. Bellot, F. Cosledan, L. Vendier, J. Brocard, B. Meunier and A. Robert, *J. Med. Chem.*, 2010, **53**, 4103.
- 44 A. Robert, O. Dechy-Cabaret, J. Cazelles and B. Meunier, *Acc. Chem. Res.*, 2002, **35**, 167; F. Benoit-Vical, J. Lelievre, A. Berry, C. Deymier, O. Dechy-Cabaret, J. Cazelles, C. Loup, A. Robert, J. F. Magnaval and B. Meunier, *Antimicrob. Agents Chemother.*, 2007, **51**, 1463.
- 45 J. J. Walsh, D. Coughlan, N. Heneghan, C. Gaynor and A. Bell, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 3599.
- 46 I. Opsenica, D. Opsenica, C. A. Lanteri, L. Anova, W. K. Milhous, K. S. Smith and B. A. Salaja, *J. Med. Chem.*, 2008, **51**, 6216.
- 47 S. Gemma, G. Campiani, S. Butini, B. P. Joshi, G. Kukreja, S. S. Coccone, M. Burrutti, M. Persico, V. Nacci, I. Fiorini, E. Novellino, D. Taramerlli, N. Banilico, S. Parapini, V. Yardley, S. Croft, S. K. Maerk, M. Rottman, R. Brun, M. Coletta, S. Marini, G. Guiso, S. Caccia and C. Fattorusso, *J. Med. Chem.*, 2009, **52**, 502.
- 48 I. Chiyanzu, C. Clarkson, P. J. Smith, J. Gut, P. J. Rosenthal and K. Chibale, *Bioorg. Med. Chem.*, 2005, **13**, 3249.
- 49 C. E. Gutteridge, D. A. Nichols, S. M. Curtis, D. S. Thota, J. V. Vo, L. Gerena, G. Montip, C. O. Asher, D. S. Diaz, C. A. DiTusa, K. Smith and A. K. Bhattacharjee, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 5682.
- 50 M. Liu, P. Wilairat and M. L. Go, *J. Med. Chem.*, 2001, **44**, 4443.
- 51 M. Chen, T. G. Theander, S. B. Christensen, L. Hviid, L. Zhai and A. Kharazmi, *Antimicrob. Agents Chemother.*, 1994, **38**, 1470.
- 52 M. Chen, S. B. Christensen, L. Zhai, M. H. Rasmussen, T. G. Theander, S. Frøkjaer, B. Steffansen, J. Davidsen and A. Kharazmi, *J. Infect. Dis.*, 1997, **176**, 1327.
- 53 N. Sriwilaijaroen, M. Liu, M. L. Go and P. Wilairat, *Southeast Asian J. Trop. Med. Public Health*, 2006, **37**, 607.
- 54 J. N. Dominguez, J. E. Charris, G. Lobo, N. G. de Dominguez, M. M. Moreno, F. Riggione, E. Sanchez, J. Olson and P. J. Rosenthal, *Eur. J. Med. Chem.*, 2001, **36**, 555.
- 55 M. L. Go, M. Liu, P. Wilairat, P. J. Rosenthal, K. J. Saliba and K. Kirk, *Antimicrob. Agents Chemother.*, 2004, **48**, 3241.
- 56 E. M. Guantai, K. Ncokazi, T. J. Egan, J. Gut, P. J. Rosenthal, P. J. Smith and K. Chibale, *Bioorg. Med. Chem.*, 2010, **18**, 8243.
- 57 K. V. Sashidhara, M. Kumar, R. K. Modukuri, R. K. Srivastava, A. Soni, K. Srivastava, S. V. Singh, J. K. Saxena, H. M. Gauniyal and S. K. Puri, *Bioorg. Med. Chem.*, 2012, **20**, 2971.
- 58 K. V. Sashidhara, S. R. Avula, G. R. Palnati, S. V. Singh, K. Srivastava, S. K. Puri and J. K. Saxena, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 5455.
- 59 B. Insuasty, A. Montoya, D. Becerra, J. Quiroga, R. Abonia, S. Robledo, I. D. Vélez, Y. Upegui, M. Nogueras and J. Cobo, *Eur. J. Med. Chem.*, 2013, **67**, 252.
- 60 F. J. Smit and D. D. N'Da, *Bioorg. Med. Chem.*, 2014, **22**, 1128.
- 61 S. Manohar, U. C. Rajesh, S. I. Khan, B. L. Tekwani and D. S. Rawat, *ACS Med. Chem. Lett.*, 2012, **3**, 555.
- 62 K. Singh, H. Kaur, K. Chibale, J. Balzarini, S. Little and P. V. Bharatam, *Eur. J. Med. Chem.*, 2012, **52**, 82.
- 63 S. I. Pretorius, W. J. Breytenbach, C. de Kock, P. J. Smith and D. D. N'Da, *Bioorg. Med. Chem.*, 2012, **21**, 269.
- 64 A. Thakur, S. I. Khan and D. S. Rawat, *RSC Adv.*, 2014, **4**, 20729.
- 65 D. Kumar, S. I. Khan, B. L. Tekwani, P. Ponnann and D. S. Rawat, *Eur. J. Med. Chem.*, 2015, **89**, 490.

- 66 S. Manohar, S. I. Khan and D. S. Rawat, *Chem. Biol. Drug Des.*, 2011, **78**, 124.
- 67 A. Kumar, K. Srivastava, S. R. Kumar, M. I. Siddiqi, S. K. Puri, J. K. Sexana and P. M. S. Chauhan, *Eur. J. Med. Chem.*, 2011, **46**, 676.
- 68 H. R. Bhat, U. P. Singh, P. Gahtori, S. K. Ghosh, K. Gogoi, A. Prakash and R. K. Singh, *New J. Chem.*, 2013, **37**, 2654.
- 69 H. R. Bhat, U. P. Singh, P. Gahtori, S. K. Ghosh, K. Gogoi, A. Prakash and R. K. Singh, *RSC Adv.*, 2013, **3**, 2942.
- 70 F. A. Rojas Ruiz, R. N. García-Sánchez, S. V. Estupiñan, A. Gómez-Barrio, D. F. Torres Amado, B. M. Pérez-Solórzano, J. J. Nogal-Ruiz, A. R. Martínez-Fernández and V. V. Kouznetsov, *Bioorg. Med. Chem.*, 2011, **19**, 4562.
- 71 K. Chauhan, M. Sharma, J. Saxena, S. V. Singh, P. Trivedi, K. Srivastava, S. K. Puri, J. K. Saxena, V. Chaturvedi and P. M. S. Chauhan, *Eur. J. Med. Chem.*, 2013, **62**, 693.
- 72 V. R. Solomon, W. Haq, K. Srivastava, S. K. Puri and S. B. Katti, *J. Med. Chem.*, 2007, **50**, 394.
- 73 S. Gemma, G. Campiani, S. Butini, G. Kukreja, S. S. Coccone, B. P. Joshi, M. Persico, V. Nacci, I. Fiorini, E. Novellino, E. Fattorusso, O. Tagliatalata-Scafati, L. Savini, D. Taramelli, N. Basilico, S. Parapini, G. Morace, V. Yardley, S. Croft, M. Coletta, S. Marini and C. Fattorusso, *J. Med. Chem.*, 2008, **51**, 1278.
- 74 S. Gemma, C. Camodeca, S. S. Coccone, B. P. Joshi, M. Bernetti, V. Moretti, S. Brogi, M. C. B. de Macros, L. Savini, D. Taramelli, N. Basilico, S. Parapini, M. Rottmann, R. Brun, S. Lamponi, S. Caccia, G. Guiso, R. L. Summers, R. E. Martin, S. Saponara, B. Gorelli, E. Novellino, G. Campiani and S. Butini, *J. Med. Chem.*, 2012, **55**, 6948.
- 75 C. Ohrt, G. D. Willingmyre, P. Lee, C. Knirsch and W. Milhous, *Antimicrob. Agents Chemother.*, 2002, **46**, 2518.
- 76 A. E. Yeo and K. H. Rieckmann, *Int. J. Parasitol.*, 1995, **25**, 531.
- 77 A. B. Sidhu, Q. Sun, L. J. Nkrumah, M. W. Dunne, J. C. Sacchettini and D. A. Fidock, *J. Biol. Chem.*, 2007, **282**, 2494.
- 78 M. Perić, A. Fajdetić, R. Rupčić, S. Alihodžić, D. Zihner, M. Bukvić Krajačić, K. S. Smith, Z. Ivezić-Schönfeld, J. Padovan, G. Landek, D. Jelić, A. Hutinec, M. Mesić, A. Ager, W. Y. Ellis, W. K. Milhous, C. Ohrt and R. Spaventi, *J. Med. Chem.*, 2012, **55**, 1389.
- 79 D. Pesic, K. Starcevic, A. Toplak, E. Herreros, J. Vidal, M. J. Almela, D. Jelic, S. Alihodzic, R. Spaventi and M. Peric, *J. Med. Chem.*, 2012, **55**, 3216.
- 80 P. Singh, P. Singh, M. Kumar, J. Gut, P. J. Rosenthal, K. Kumar, V. Kumar, M. P. Mahajan and K. Bisetty, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 57.
- 81 R. Raj, C. Biot, S. Carrère-Kremer, L. Kremer, Y. Guérardel, J. Gut, P. J. Rosenthal and V. Kumar, *Chem. Biol. Drug Des.*, 2014, **83**, 191.
- 82 P. Singh, R. Raj, P. Singh, J. Gut, P. J. Rosenthal and V. Kumar, *Eur. J. Med. Chem.*, 2014, **71**, 128.
- 83 R. Raj, V. Mehra, J. Gut, P. J. Rosenthal, K. J. Wicht, T. J. Egan, M. Hopper, L. A. Wrischnik, K. M. Land and V. Kumar, *Eur. J. Med. Chem.*, 2014, **84**, 425.
- 84 S. P. Kumar, J. Gut, R. C. Guedes, P. J. Rosenthal, M. M. M. Santos and R. Moreira, *Eur. J. Med. Chem.*, 2011, **46**, 927.
- 85 R. Raj, P. Singh, P. Singh, J. Gut, P. J. Rosenthal and V. Kumar, *Eur. J. Med. Chem.*, 2013, **62**, 590.
- 86 R. Raj, J. Gut, P. J. Rosenthal and V. Kumar, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 756.
- 87 R. Raj, C. Biot, S. Carrère-Kremer, L. Kremer, Y. Guérardel, J. Gut, P. J. Rosenthal, D. Forge and V. Kumar, *Chem. Biol. Drug Des.*, 2014, **83**, 622.
- 88 Nisha, J. Gut, P. J. Rosenthal and V. Kumar, *Eur. J. Med. Chem.*, 2014, **84**, 566.
- 89 G. Gasser and N. Metzler-Nolte, *Curr. Opin. Chem. Biol.*, 2012, **16**, 84.
- 90 C. Biot, W. Castro, C. Y. Botte and M. Navarro, *Dalton Trans.*, 2012, **41**, 6335; C. G. Hartinger and P. J. Dyson, *Chem. Soc. Rev.*, 2009, **38**, 391; G. Jaouen, *Bioorganometallics: Biomolecules, Labeling, Medicine*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, 2006.
- 91 M. A. Jakupec, M. Galanski, V. B. Arion, C. G. Hartinger and B. K. Keppler, *Dalton Trans.*, 2008, 183.
- 92 J. A. Ocheskey, S. E. Harpstrite, A. Oksman, D. E. Goldberg and V. Sharma, *Chem. Commun.*, 2005, **12**, 1622.
- 93 C. Biot, G. Glorian, L. A. Maciejewski, J. S. Brocard, O. Domarle, G. Blampain, P. Millet, A. J. Georges, H. Abessolo, D. Dive and J. Lebib, *J. Med. Chem.*, 1997, **40**, 3715.
- 94 N. Chavain, E. Davioud-Charvet, X. Trivelli, L. Mbeki, M. Rottmann, R. Brun and C. Biot, *Bioorg. Med. Chem.*, 2009, **17**, 8048.
- 95 R. Arancibia, F. Dubar, B. Pradines, I. Forfar, D. Dive, A. H. Klahn and C. Biot, *Bioorg. Med. Chem.*, 2010, **18**, 8085.
- 96 P. F. Salas, C. Herrmann, J. F. Cawthray, C. Nimphius, A. Kenkel, J. Chen, C. de Kock, P. J. Smith, B. O. Patrick, M. J. Adam and C. Orvig, *J. Med. Chem.*, 2013, **56**, 1596.
- 97 C. Herrmann, P. F. Salas, J. F. Cawthray, C. de Kock, B. O. Patrick, P. J. Smith, M. J. Adam and C. Orvig, *Organometallics*, 2012, **31**, 5736.
- 98 F. Bellot, F. Cosledan, L. Vendier, J. Brocard, B. Menunier and A. Robert, *J. Med. Chem.*, 2010, **53**, 4103.
- 99 L. Glans, D. Taylor, C. de Kock, P. J. Smith, M. Haukka, J. R. Moss and E. Nordlander, *J. Inorg. Biochem.*, 2011, **105**, 985.
- 100 S. D. Khanye, J. Gut, P. J. Rosenthal, K. Chibale and G. S. Smith, *J. Organomet. Chem.*, 2011, **696**, 3296.
- 101 L. Glans, W. Hu, C. Jost, C. de Kock, P. J. Smith, M. Haukka, H. Bruhn, U. Schatzschneider and E. Nordlander, *Dalton Trans.*, 2012, **41**, 6443.
- 102 Y. Li, C. de Kock, P. J. Smith, K. Chibale and G. S. Smith, *Organometallics*, 2014, **33**, 4345.
- 103 K. Kumar, S. Schniper, A. G. Alez-Sarrias, A. A. Holder, N. Sanders, D. Sullivan, W. L. Jarrett, K. Davis, F. Bai, N. P. Seeram and V. Kumar, *Eur. J. Med. Chem.*, 2014, **86**, 81.
- 104 D. De, F. M. Krogstad, L. D. Byers and D. J. Krogstad, *J. Med. Chem.*, 1998, **41**, 4918.

- 105 T. S. Feng, E. M. Guantai, M. J. Nell, C. E. J. van Rensburg, H. C. Hoppe and K. Chibale, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 2882.
- 106 M. C. Lombard, D. D. N'Da, J. C. Breytenbach, P. J. Smith and C. A. Lategan, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 6975.
- 107 M. C. Lombard, D. D. N'Da, J. C. Breytenbach, P. J. Smith and C. A. Lategan, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 1683.
- 108 T. S. Feng, E. M. Guantai, M. J. Nell, E. J. Constance, V. Rensburg, K. Ncokazi, T. J. Egan, H. C. Hoppe and K. Chibale, *Biochem. Pharmacol.*, 2011, **82**, 236.
- 109 F. Coslédan, L. Fraisse, A. Pellet, F. Guillou, B. Mordmüller, P. G. Kremsner, A. Moreno, D. Mazier, J. P. Maffrand and B. Meunier, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 17579.
- 110 N. C. P. Araújo, V. Barton, M. Jones, P. A. Stocks, S. A. Ward, J. Davies, P. G. Bray, A. E. Shone, M. L. S. Cristiano and P. M. O'Neill, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 2038.
- 111 L. Gupta, K. Srivastava, S. Singh, S. K. Puri and P. M. S. Chauhan, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 3306.
- 112 V. K. Zishiri, M. C. Joshi, R. Hunter, K. Chibale, P. J. Smith, R. L. Summers, R. E. Martin and T. J. Egan, *J. Med. Chem.*, 2011, **54**, 6956.
- 113 F. Kobarfard, V. Yardley, S. Little, F. Daryaei and K. Chibale, *Chem. Biol. Drug Des.*, 2012, **79**, 326.
- 114 A. Kumar, K. Srivastava, S. R. Kumar, S. K. Puri and P. M. S. Chauhan, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 7059.
- 115 C. C. Musonda, G. A. Whitlock, M. J. Witty, R. Brun and M. Kaiser, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 481.
- 116 C. J. A. Ribeiro, S. P. Kumar, J. Gut, L. M. Gonçalves, P. J. Rosenthal, R. Moreira and M. M. M. Santos, *Eur. J. Med. Chem.*, 2013, **69**, 365.
- 117 K. Ersmark, B. Samuelsson and A. Hallberg, *Med. Res. Rev.*, 2006, **26**, 626.
- 118 N. Vaiana, M. Marzahn, S. Parapini, P. Liu, M. Dell'Agli, A. Pancotti, E. Sangiovanni, N. Basilico, E. Bosisio, B. M. Dunn, D. Taramelli and S. Romeo, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 5915.
- 119 M. Mungthin, P. G. Bray, R. G. Ridley and S. A. Ward, *Antimicrob. Agents Chemother.*, 1998, **42**, 2973.
- 120 W. A. Andayi, T. J. Egan, J. Gut, P. J. Rosenthal and K. Chibale, *ACS Med. Chem. Lett.*, 2013, **4**, 642.
- 121 L. van Heerden, J. C. Breytenbach, C. de Kock, P. J. Smith, J. W. Breytenbach, T. T. Cloete and D. D. N'Da, *Eur. J. Med. Chem.*, 2012, **55**, 335.
- 122 M. N. Aminake, A. Mahajan, V. Kumar, R. Hans, L. Wiesner, D. Taylor, C. de Kock, A. Grobler, P. J. Smith, M. Kirschner, A. Rethwilm, G. Pradel and K. Chibale, *Bioorg. Med. Chem.*, 2012, **20**, 5277.
- 123 A. Parthiban, J. Muthukumaran, A. Manhas, K. Srivastava, R. Krishna and H. S. P. Rao, *Bioorg. Med. Chem. Lett.*, 2015, DOI: 10.1016/j.bmcl.2015.08.030.
- 124 D. S. B. Ongarora, N. Strydom, K. Wicht, M. Njoroge, L. Wiesner, T. J. Egan, S. Wittlin, U. Jurva, C. M. Masimirembwa and K. Chibale, *Bioorg. Med. Chem.*, 2015, **23**, 5419.
- 125 A. H. Romero, M. E. Acosta, N. Gamboa, J. E. Charris, J. Salazar and S. E. Lopez, *Bioorg. Med. Chem.*, 2015, **23**, 4755.